Coprinopsis natarajanii sp. nov., a new marine basidiomycete from Puducherry mangroves, East coast of India

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ABSTRACT

Only a few basidiomycetous marine fungi are known from marine environments, while *Ascomycota* predominate the marine environments. In this paper, a novel species of *Coprinopsis* is reported from Indian mangroves based on the morphological and molecular phylogenetic analyses. The basidiomata and the pleurocystidia of the new species are smaller when compared to the other species of *Coprinopsis*.

Keywords: Basidiomycete, Mangrove fungi, Novel species, Phylogeny, Taxonomy

INTRODUCTION

Jones et al. (2019) listed 1,257 species of marine fungi belonging to 539 genera of which only 22 species in 17 genera belong to filamentous Basidiomycota whereas 805 species in 352 genera belong to Ascomycota. Devadatha et al. (2021) reported 850 mangrove fungi of which 58 species belong to Basidiomycota. Sarma and Devadatha (2020) listed 35 species in 28 genera from Basidiomycota reported from Indian mangroves including the terrestrial basidiomycetes. Marine Basidiomycetes are few in number when compared to their terrestrial counterparts and are an ecological group, and taxonomically diverse (Agariomycotina, Uredinomycotina and Ustilaginomycotina). Marine Basidiomycetes are well adapted to their habitats, with reduced basidiomata. Marine species are known only as teleomorphs with basidiospores generally released passively (Jones and Choeyklin 2008). Jones (2011) indicated that many Basidiomycetes are still to be discovered and described.

Studies on the macrofungal diversity in mangroves are scant and this is especially true with respect to India. Dutta et al. (2013) and Ghate and Sridhar (2016) have given an insight into the macrofungal diversity associated with mangroves of Sunderbans and Southwest coast of India, respectively. A number of studies on microfungal diversity from mangrove plants or habitats have been carried out in India (Borse 1988; Chinnaraj 1993; Ravikumar and Vittal 1996; Sarma and Vittal 2000, 2001; Maria and Sridhar 2002, 2003, 2004; Vittal and Sarma 2006). In a survey on litter degrading fungi from Puducherry mangroves a species of Coprinopsis P. Karst. was recorded. This species of Coprinopsis, was found to be a novel species although 27 species of Coprinopsis have been reported from India (Amandeep et al. 2014). Coprinopsis is characterized by cutis type pileipellis that differentiates it from its related genera like Coprinellus P. Karst., and Parasola Redhead, Vilgalys & Hopple., in Psathyrellaceae (Readhead et al. 2001). Further, the presence of pleurocystidia differentiates it from Coprinus Pers. that is presently placed in the family Agaricaceae. Amandeep et al. (2014) documented the diversity of *Coprinopsis* species from the coprophilous habitats in India.

Coprinopsis recorded in the present study was studied

morphologically. However, due to absence of mature specimens, the description of the species lacks details of basidiospores and basidia. Since there are no reports of occurrence of *Coprinopsis* in the submerged parts of the mangroves, ITS sequence analysis was done to prove the novelty of the species.

MATERIALS AND METHODS

Morphological characterization

The dead and decaying samples of Avicennia marina (Forssk.) Vierh. wood was collected in large plastic bags from Pondicherry mangroves, Puducherry in India and transported to the laboratory. Morphological characters were studied using an Optika stereo zoom SZM-LED1 microscope and Nikon ECLIPSE TiU upright microscope, photographs were taken using Nikon DS-Fi2 digital camera fitted to the microscope. Microscopic examination of anatomical characters was done following Largent et al. (1977). Photoplates were prepared using Adobe Photoshop CS6 updated version 13.0.1 software (Adobe Systems Inc., The United States) while Nikon NIS-Elements-Imaging Software version 4.4 program was used for measurements. The specimen (air-dried wood with fungal material) was deposited at Ajrekar Mycological Herbarium (AMH), Agharkar Research Institute (ARI), Pune, India.

DNA extraction and PCR amplification

Genomic DNA was extracted using the GeneJET Plant Genomic DNA Purification kit (Thermo Fisher Scientific Inc., US) following the manufacturer's protocol, from the freshly collected specimens of *Coprinopsis* species. DNA purity was checked and estimated by Thermo ScientificTM NanoDrop 2000. In this study, the ITS sequence was amplified by PCR, following methods laid down by White *et al.* (1990). All PCR amplification reactions were performed in an Eppendorf Master cycler with a total volume of 50μL, consisting of 25μL of AMPLIQON Taq DNA Polymerase 2X Master Mix RED, 1 μL of each primer (10 μM), 2μL DNA template and the remaining volume was made up with that of HiMEDIA Molecular Biology Grade Water.

The PCR amplification products were checked on 1% agarose gel stained with ethidium bromide. The PCR products were

purified by using QIAquick PCR Purification Kit (QIAGEN, Germany) by following manufacturer's protocol. The purified PCR products were estimated by Thermo ScientificTM NanoDrop 2000 and DNA sequencing was outsourced.

Sequence alignment and molecular phylogeny

Sequences used in the phylogenetic analyses were downloaded from GenBank based on BLAST search similarity and recently published data (Gierczyk *et al.* 2017; Nguyen *et al.* 2019). Sequence alignments were performed for individual gene regions at (http://mafft.cbrc.jp/alignment/server), using iterative refinements as E-INS-i method for ITS. The unaligned portions were aligned manually using BioEdit v.7.0.5.2 (Hall 1999). The sequence alignment was converted to PHYLIP format (.phy) using ALTER (alignment transformation environment: (http://sing.eiuvigo.es/ALTER/, 2017)) for Randomized Accelerated Maximum Likelihood (RAxML) analysis.

Maximum Likelihood analyses were conducted using the RAxML-HPC2 on XSEDE (8.2.8) with GTR + G + I model (Stamatakis *et al.* 2008, Stamatakis 2014) in the CIPRES Science Gateway platform (Miller *et al.* 2010). Phylogenetic trees were visualized with FigTree v1.4.0 program (Rambaut 2012) and reorganized in Microsoft power point (2016). Sequences generated in this study were deposited in GenBank.

RESULTS

Taxonomy

Coprinopsis natarajanii Devadatha, V. Kumaresan and V.V. Sarma sp. nov.

Holotype: AMH-10164

Index Fungorum No.: IF558487; Figs. a-f

Etymology: In honour of Late Prof. K. Natarajan of CAS in Botany, University of Madras for his contributions to *Basidiomycota* of India.

Basidiomata gregarious (Fig. 1a). Pileus 2-10 mm diam. in button stage with closed pileus, 6-18 mm diam, when mature, conical to campanulate, white to pale brown at the disc, white towards the margin, pileal veil appressed fibrillose, flesh thin, membranous. Lamellae free to adnexed, crowded, up to 0.1 cm broad, white, concolorous. Stipe 10-20 × 3-5 mm, cylindrical, slightly tapering upwards, white, covered with fine erect fibrils, sericeous underneath, context hollow, white. Basidia and basidiospores not observed. Basidioles clavate, $18-21 \times 9-11$ μm. Pleurocystidia 50-60 μm × 10-15 μm, elongate clavate to fusoid ventricose. Cheilocystidia 40-50 μm × 6-10 μm, similar in shape to pleurocystidia. Pileus cuticle a cutis having veil in the form of hyphal chains. Context thin, composed of filamentous, radially arranged thin-walled hyphae, 4-10 µm, Hymenophoral trama subregular, composed of thin-walled, hyaline, hyphae up to 11 µm broad. Stipe cuticle hyphal, smooth, context white. Clamp connections observed. Smells mushroomy, no taste recorded.

Material examined: INDIA, Tamil Nadu, Pondicherry,

Veerampattinam mangroves, on decaying wood of *Avicennia marina* (*Acanthaceae*), 20 January 2017, B. Devadatha (AMH-10164, holotype).

Notes: Morphological details and the BLAST search analysis based on ITS sequence data showed that the collected taxon belongs to the genus Coprinopsis. Further, phylogenetic analysis along with ITS sequence data of Coprinopsis species revealed that the present new taxon, Coprinopsis natarajanii, is clustered in the clade containing Coprinopsis taxa with strong support from maximum likelihood analyses (98% BS). The morophology of C. natarajanii is distinct from C. phaeopunctata (Esteve-Ray. & A. Ortega) Valade in having smaller size of pleurocystidia (50-60 μ m × 10-15 μ m vs 35-75 $(100) \times 25-50 \mu m$). The new species described herein was recorded on decaying wood of Avicennia marina, a mangrove plant in marine environments, whereas C. phaeopunctata has been recorded on rotten wood and debris of Abies pinsapo (Ortega and Esteve-Raventos 2003) from terrestrial environments. Further, ITS nucleotide sequence comparison between C. phaeopunctata and C. natarajanii revealed 4.2% (28) nucleotide base pair difference which supports the establishment of a new species for our new taxon (Jeewon and Hyde, 2017). Thus, based on the differences in the molecular analysis as shown in the phylogenetic tree and the differences in the micromorphological characters, substrate and habitat, the new species, C. natarajanii is introduced in the genus Coprinopsis.

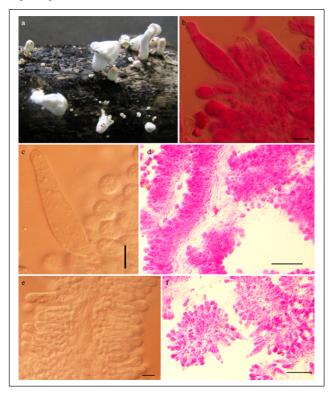


Fig. 1: Coprinopsis natarajanii a-b. Basidiomata on the decaying wood. b-c. Pleurocystidia. d-e. Basidioles and pseudoparaphysis in gill trama. f. Cheilocystidia. Scale bars: $d_1f = 50 \mu m$. b-c, $e = 10 \mu m$

Molecular phylogeny

The ITS gene dataset composed of 69 taxa with 763 nucleotide characters from *Psathyrellaceae*, with *Lacrymaria lacrymabunda* SZMC:NL:2140 as outgroup. RAxML analysis of the ITS gene dataset yielded the best scoring tree (Fig. 1) with a final ML optimization likelihood value of 7635.776556. The matrix had 464 distinct alignment patterns, with 21.82% of undetermined characters or gaps. Estimated base frequencies were as follows: A = 0.233036, C = 0.236203, G = 0.230836, and T = 0.299925; substitution rates AC = 1.386642, AG = 2.956964, AT = 1.907401, CG = 0.514276, CT = 4.810990, and GT = 1.000000; proportion of invariable sites I = 0.320066; and gamma distribution shape parameter α = 0.592406.

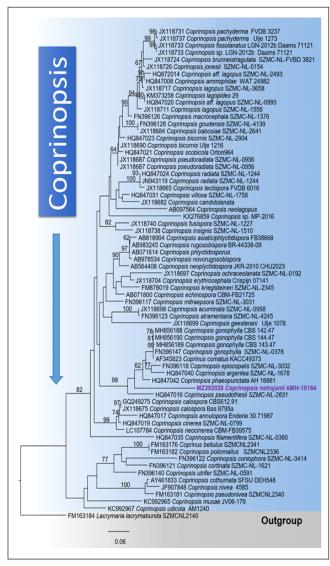


Fig. 2: Phylogram generated through maximum likelihood analyses based on nrITS nucleotide sequence data for *Coprinopsis natarajanii* and other species of *Coprinopsis* with *Lacrymaria lacrymabunda* as the outgroup taxon. Maximum likelihood bootstrap values ≥60% are given at the nodes. The new isolate is in purple colour

Phylogenetic analyses based on ITS nucleotide sequence dataset indicate that our new species Coprinopsis natarajanii grouped within the Coprinopsis. Our taxon C. natarajanii forms a distinct lineage basal to Coprinopsis phaeopunctata although, the position is unsupported. The tree topology is supported by lower bootstrap values (62% ML; Fig. 2) indicating that C. natarajanii is a new taxon.

DISCUSSION

Only a few filamentous *Basidiomycetes* are considered as marine fungi reported from marine environments. In our ongoing investigations on the marine fungal diversity in mangroves of Pondicherry, East coast of India, we have encountered a small basidiomycete which fits in *Coprinopsis*. Further characterisation of the taxon based on morphological and molecular phylogenetic analyses, based on single-locus (ITS) maximum likelihood tree revealed that it is a novel species within the genus *Coprinopsis*. Hence *C. natarajanii* is described herein as a new species.

Coprinopsis species are normally recorded from dung habitats (Amandeep et al., 2014) and very few from plant litter. A few species of Coprinopsis have earlier been reported from mangroves including Coprinopsis atramentaria (Bull.) Redhead, Vilgalys & Moncalvo and C. lagopus (Fr.) Redhead, Vilgalys & Moncalvo, but not as marine fungi. Coprinopsis lagopus has been recorded from Sundarban mangroves (Dutta et al. 2013), which is known to possess larger fruit-bodies (Pileus with 30-90 mm diam.) and broader pleuro- and cheilocystidia compared to C. natarajanii. Li et al. (2016), while studying the endophyte diversity of mangrove fungi from southern China, isolated Coprinopsis atramentaria from the twigs of Kandelia candel. The identification of this species, however, was based on ITS sequence analysis. Coprinopsis atramentaria has earlier been described to consist larger fruit-bodies (Pileus with 30-60 mm diam) and longer and broader pleuro- and cheilocystidia (Orton and Watling 1979).

Molecular studies revealed, the genus *Coprinus* is subdivided into *Coprinus sensu stricto* (*Agaricaceae*), *Coprinellus*, *Coprinopsis* and *Parasola* in the family *Psathyrellaceae* and among these the generic name *Coprinopsis* P. Karst. has not been in active use for 100 years (Redhead *et al.* 2001). It is characterized by pileipellis, which is not a hymeniderm, not a cystoderm but a cutis. At present there are about 150 species belonging to this genus. Most of them are known to colonize dung materials. Very few are known from other substrata.

Our new species, as already mentioned in the notes section, differs morphologically and also on molecular basis. Hence the new species *C. natarajanii* in the genus *Coprinopsis* is reported in this paper.

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REFERENCES

- Amandeep, K., Atri, N.S. and Munruchi, K., 2014. Taxonomic study on coprophilous species of *Coprinopsis (Psathyrellaceae, Agaricales)* from Punjab, India. *Mycosphere* **5(1)**: 1-25.
- Borse, B.D. 1988. Frequency of occurrence of marine fungi from Maharashtra coast, India. *Indian J. Geo-Marine Sci.* **17**: 165-167.
- Chinnaraj, S. 1993. Higher marine fungi from mangroves of Andaman and Nicobar Islands. *Sydowia* **45**: 109-115
- Devadatha, B., Jones, E.B.G., Pang, K.L., Abdel-Wahab, M.A., Hyde, K.D., Sakayaroj, J., Bahkali, A.H., Calabon, M.S., Sarma, V.V., Sutreong, S. and Zhang, S.N., 2021. Occurrence and geographical distribution of mangrove fungi. *Fungal Diversity* **106 (1)**:137-227.
- Dutta, A.K., Pradhan, P., Basu, S.K. and Acharya, K., 2013. Macrofungal diversity and ecology of the mangrove ecosystem in the Indian part of Sundarbans. *Biodiversity* **14 (4)**:196-206.
- Ghate, S.D. and Sridhar, K.R., 2016. Contribution to the knowledge on macrofungi in mangroves of the Southwest India. *Plant Biosystems* **150 (5)** : 977-986.
- Gierczyk, B., Kujawa, A. and Szczepkowski, A. 2014. New to Poland species of the broadly defined genus *Coprinus (Basidiomycota, Agaricomycotina)*. *Acta Mycol.* 49 (2): 159-188.
- Hall, T.A., 1999, January. BioEdit: A User-friendly Biological Sequence Alignment Editor and Analysis Program for Windows 95/98/NT. <u>In: Nucleic acids symposium series 41</u>: 95-98.
- Jeewon, R. and Hyde, K.D. 2016. Establishing species boundaries and new taxa among fungi: recommendations to resolve taxonomic ambiguities. *Mycosphere* 7: 1669-1677.
- Jones, E.B.G., and Choeyklin, R., 2008. Ecology of Marine and Freshwater *Basidiomycetes*. In: *British Mycological Society Symposia Series* **28**: 301-324. Academic Press.
- Jones, E.B.G., 2011. Are there more marine fungi to be described? *Bot. Mar.* **54**: 343-354.
- Jones, E.B.G., Pang, K.L., Abdel-Wahab, M.A., Scholz, B., Hyde, K.D., Boekhout, T., Ebel, R., Rateb, M.E., Henderson, L., Sakayaroj, J. and Suetrong, S. 2019. An online resource for marine fungi. *Fungal*

- Diversity 96(1):347-433.
- Largent, D.L., Johnson, D. and Watling, R. 1977. How to Identify Mushrooms to Genus III: Microscopic features, Mad River Press Inc., California.
- Li, J.L., Sun, X., Chen, L. and Guo, L.D. 2016. Community structure of endophytic fungi of four mangrove species in Southern China, *Mycology* **7 (4)**: 180-190.
- Maria, G.L. and Sridhar, K.R. 2002. Richness and diversity of filamentous fungi on woody litter of mangroves along the west coast of India. *Curr. Sci.* **83** (12): 1573-1580.
- Maria, G.L. and Sridhar, K.R. 2003. Diversity of filamentous fungi on woody litter of five mangrove plant species from the southwest coast of India. *Fungal Diversity* **14**: 109-126.
- Maria, G.L. and Sridhar, K.R. 2004. Fungal colonization of immersed wood in mangroves of the southwest coast of India. *Can. J. Bot.* **82**: 1409-1418.
- Miller, M.A., Pfeiffer, W. and Schwartz, T., 2012. The CIPRES Science Gateway: Enabling High-impact Science for Phylogenetics Researchers with Llimited Resources. In: Proceedings of the 1st Conference of the Extreme Science and Engineering Discovery Environment: Bridging from the Extreme to the Campus and Beyond, 18.
- Nguyen, P.T., Pham, N.D.H., Suzuki, A., Shimizu, K. and Fukiharu, T., 2019. *Coprinopsis neocinerea* sp. nov., an ammonia fungus from Southern Vietnam. *Mycoscience* **60 (5)**: 307-312.
- Ortega, A. and Esteve-Raventós, F. 2005. A new species of *Gymnopilus (Cortinariaceae)* from sandy soils in *Pinus* forests. *Persoonia* **18**: 505-10.
- Orton, P.D. and Watling, R. 1979. British Fungus Flora, Part 2: *Coprinaceae*: *Coprinus*. Royal Botanic Gardens, Edinburgh. pp.149.
- Rambaut, A., 2012. FigTree v1. 4. Available online at: http://tree.bio.ed.ac.uk/software/figtree
- Ravikumar, D.R. and Vittal, B.P.R. 1996. Biodiversity of mangrove fungi on different substrata of *Rhizophora apiculata* and *Avicennia* spp. from Godavari and Krishna deltas, east coast of India. *Indian J. Geo-Marine Sci.* 2:142-144.
- Redhead, S.A., Vilgalys, R., Moncalvo, J.M., Johnson, J. and Hopple Jr, J.S., 2001. *Coprinus* Pers. and the disposition of *Coprinus* species *sensu lato*. *Taxon* **50 (1)**: 203-241.
- Sarma, V.V.; Devadatha, B. 2020. Fungal Diversity in Mangroves of India and a Note on their Medicinal Potential. In: *Biotechnological Utilization of Mangrove Resources*. (Eds.: Patra J.K., Mishra R.R.

- and Thatoi, H.) Elsevier, pp. 153-224.
- Sarma, V.V. and Vittal, B.P.R. 2000. Biodiversity of mangrove fungi on different substrata of *Rhizophora apiculata* and *Avicennia* spp. from Godavari and Krishna deltas, east coast of India. *Fungal Divers.* 5: 23-41.
- Sarma, V.V. and Vittal, B.P.R. 2001. Biodiversity of manglicolous fungi on selected plants in the Godavari and Krishna deltas, East coast of India. *Fungal Divers.* **6**:115-130.
- Stamatakis, A., Hoover, P. and Rougemont, J., 2008. A rapid bootstrap algorithm for the RAxML web servers. *Systematic biology* **57 (5)**: 758-771.

- Stamatakis, A., 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**(**9**):1312-1313.
- Vittal, B.P.R. and Sarma, V.V. 2006. Diversity and ecology of fungi on mangroves of Bay of Bengal region An overview. *Indian J. Geo-Marine Sci.* **35 (4)** : 308-317.
- White, T.J., Bruns, T., Lee, S.J.W.T. and Taylor, J., 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. PCR protocols: a guide to methods and applications 18 (1): 315-322.