

Nutraceutical, pharmaceuticals and industrial bioactive compounds of gasteroid fungi: A review

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(Submitted on September 16, 2021; Accepted on November 08, 2021)

ABSTRACT

The gasteroid fungi are the group of macro-fungi within the division *Basidiomycota* which produces spores inside the basidium unlike the other members of basidiomycetous mushroom. The gasteroid fungi are well recognized for their nutritional as well as therapeutic values throughout the world. Their fruiting bodies have been evaluated to be a magnificent source of digestible carbohydrates, proteins and fats with essential amino acids, dietary fibers, vitamins and minerals. Therapeutic significance of this group of fungi includes anti-cancerous activity either by suppressing the tumor cell lines or due to their immuno-modulatory properties through the production of bioactive compounds. The metabolites from them have also been proved to be a natural anti-inflammatory, antibiotics and antifungal agents that may substitute the drugs to provide a healthy food supplement without any adverse effects. The bioactive substances from these macrofungi are also known to lower the blood glucose level by regulating level of alpha amylases and aldolase reductase. The gasteroid fungi also known for potential antioxidant properties and aphrodisiac activity. The present study aims to review the diversity of gasteroid fungi and their role in production of potential bioactive metabolites especially of nutraceutical and therapeutic importance.

Keywords: Bioactive compound, Gasteroid fungi, Nutritional value, Medicinal properties

INTRODUCTION

The fungi as potential nutrient recycler and decomposer play a significant role in ecosystem practices and also have their impact on humans and their activities (Mueller and Schmit, 2007). Among fungi, the macrofungi not only provides a diverse range of food and nutrition to mankind but also make available, innumerable raw material for industries, agriculture and pharmacy (Chang and Miles, 2004). A large number of wild mushrooms are edible but some are toxic too. Therefore, the identification and sensible utilization of these macrofungi is an important exercise. The edible mushrooms with medicinal properties have long been known by humans and were also been used by ancient civilizations as the source of valuable food and medicines as they known to have of high and low-molecular-weight bioactive compounds (Badalyan *et al.*, 2019)

The gasteroid (literally "stomach fungi") are the cluster of macro-fungi within the division *Basidiomycota*. This cluster was formerly placed in the obsolete class *Gasteromycetes* of the order *Gasteromycetales*. The cluster is, moreover, informally positioned in the taxonomic hierarchy as it includes an unnatural heterogeneous assemblage (puff balls, earth stars, stink horn and birds nest fungi) of *Basidiomycetes* having endogenous (within the basidiocarp) production of basidiospores instead of exogenous that occurs in other basidiomycetous fungi (Gogoi and Vipin, 2015). The cluster is polyphyletic displaying morphological variation as their species are not closely related to each other. The gasteroid fungi are the most evolved and display a significant morphological diversity at a glance from other groups of macrofungi. They are cosmopolitan and are more prevalent in dry and warmer parts of the world where soil temperature is high and soil fertility is minimal suggesting a hymenomycetous origin for adaptation to xerophytic condition (Smith and Weber, 1980), in contrast to some species of stinkhorns and their allies found in the wet tropic and also on temperate sand dunes. Nutritionally, most of the species are

saprobic in soil and some are growing on dung (coprophillus) and rotten wood (lignicolous), tree stumps, on decaying logs, or on the ground in the woods, hidden under leaf litter (Gastrum), city lawns, grassy open field and also hot desert (Gehlot *et al.*, 2020) while, a few have ectomycorrhizal association (Scleroderma) forming a mutually beneficial relationship with the roots of living trees (Gehlot and Singh, 2015). Further, there are two aquatic gasteroid fungi, *Niavibrissa* is marine and grows on the drift wood in the sea that is also called a marine white rot gasteroid fungus (Julich, 1981) and *Limnoperdon*, grows in marshy places (Escobar *et al.*, 1976; Hibbett and Binder, 2001). The fruiting bodies of most of gasteroid fungi are epigeal (Blackwell and Spatafora, 2004), although some forms do produce their basidiocarp beneath the surface of soil are called hypogean (*Rhizopogon*).

The gasteroid have characteristic angiocarpic (basidiocarp) fruit bodies together with aseptate halo-basidia and passive release of basidiospores (statismospores) as the basidia are not involved in the release of spore. These basidiocarp ranges from a few mm (2 mm in *Sphaerobolus*) to over one meter in diameter (*Calvatia*). The fruit bodies are stalked either with true (consists of parallel hyphae) or pseudo stem (made of spongy tissue). The inner fertile portion of basidiocarps known as gleba remain covered with sterile jacket called peridium that further consists of exoperidium, mesoperidium and endoperidium. Additionally, gleba comprises basidia and trama (sterile tissues) that may or may not include columella and capillitium. Columella are involved in conduction of nutrients whereas, capillitium helps in efficient dispersal of spores. The gleba at maturity may be dry and powdery or fleshy and slimy. The peridium may be firm and hard or thin and papery. The basidiocarp of gasteroid fungi generally remain permanently closed and the dispersal of spores takes place either through the disintegration of peridium through external agents or through inbuilt mechanism occurs once the spores get matured. Only a few species have been cultured in laboratory, and in most cases even the spore germination fails

on synthetic media (Phutela *et al.*, 1998; Doshi *et al.*, 1999).

A wide range of basidiomata structure occurs among the gastroid fungi and these characteristic life-forms results in the designation of many genera monotypic or having limited number of species. Trierveiler-Pereira and Baseia (2009) have recorded about 232 gasteroid species from Brazil which are distributed among the 54 genera and 16 families with few prevailing species as *Morganella fuliginea*, *Calvatia cyathiformis*, *Geastrum saccatum*, *Scleroderma albidum* and *S. verrucosomas*.

The gasteroid genus like *Scleroderma* that contains both epigeous and hypogeous components were discussed imposing the molecular techniques regarding secondary metabolites, development, spore morphology, and anatomy (Watling, 2006). The mushrooms, like higher plants, have ability of bioaccumulation of phytochemicals with potential medicinal properties (Akgul *et al.*, 2017). The synthetic drugs although provide a rapid relief but they are also toxic in many ways. Therefore, there is a need for adapting the natural pharmaceutical products with few or none adverse effects. The mushrooms exhibit antimicrobial, antibacterial, anti-carcinogenic, antioxidant, antiviral, anti-inflammatory, anticoagulant, cytotoxic, cytostatic, antiatherogenic, antioxidant, anti-allergic, hypoglycemic and immunosuppressive properties (Lam *et al.*, 2001; Sano *et al.*, 2002; Barros *et al.*, 2007; Olennikov *et al.*, 2011). In mushrooms both the fruiting bodies and mycelial mass constitutes diverse forms of compounds including flavonoids, alkaloids, polysaccharides, polyglucans, polyphenol, steroids, terpenoids, polyketides and dietary fibers with numerous pharmacological activities (Thatoi and Singdevsachan, 2014). The basidiomycetous-mushrooms are also efficient in degradation of lignin as they utilize all major cell wall components (Jasalavich *et al.*, 2000).

Determination of biological activities of mushrooms is significant to reveal new pharmacological agents. Decisively the gasteroid-mushrooms have ability for diverse therapeutic metabolites having potential biological activities and thus make it important to characterize not only their compounds but also taxonomic positions so that the researchers can screen their taxa for future biotechnological applications. The gasteroid fungi although has been exploited well for their potential use in biocontrol, bioremediation and novel bioactive compounds but extensive research is required to ensure the role these fungus for potential biotechnological and pharmacological applications. Further, out of 83 genera known of gasteroid fungi only few have been explored potentially for their bioactive compound and further, among the explored once there is not any concise report available that undertakes the nutraceutical bioactive compounds, pharmaceuticals compounds with therapeutic use like anticancerous, antibiotic, antioxidants, antidiabetic, immunomodulatory, antibacterial and antifungal properties including the industrial and ethnomycological uses and application of gasteroid bioactive compound. Therefore, this review tried the compile the scattered literature available for

majority of gasteroid genera known for significant bioactive compounds under diverse categories that likely to be beneficial for research communities.

NUTRITIONAL AND NUTRACEUTICAL BIO-ACTIVE COMPOUNDS

The edible mushrooms flourishing in their natural habitats were always conventionally valued for their health promoting attributes (Crisan and Sands, 1978) and were exploited as a prevailing nutraceutical source having great aroma and flavor (Kalač, 2016). The great amount of energy with low amount of fats makes them an important food supplement and natural endowments. The fruit bodies gasteroid mushrooms are found to be magnificent source of digestible carbohydrates and protein with essential amino acids, dietary fibers, vitamins and minerals (Yin and Zhou, 2008; Liu *et al.*, 2012; Vishwakarma *et al.*, 2017). The mushrooms in the current world are also regarded as therapeutic foods or nutraceuticals due to the presence of compound in their fruit bodies having great pharmacological properties (Barros *et al.*, 2008).

Astraeus hygrometricus is known to have too hard fruiting bodies to be consumed in North American regions (Miller and Miller, 2006). In contrast to this its collected and sold in the markets of India and Nepal as one of the delicious foods (Maiti *et al.*, 2008). In southeast Asia the species of *Astraeus* were reported to have an abundance of volatile compound of eight carbon including furfural, benzaldehyde, cyclohexenone, and furanyl (Kakumyan and Matsui, 2009). Similarly, the *Battarrea phalloides* and *B. stevenii* are also described as unknown edibility. However, in Cyprus, the immature egg-form of the fruit body is eaten (Miller and Miller, 2006; Roberts and Evans, 2011). The young fruiting bodies or the puffballs of the genus *Bovista* are edible till they are white inside as they can be jumbled with immature *Amanitas* which are potentially deadly harmful (McKnight and McKnight, 1998). Additionally, the immature fruit bodies of *Geastrum triplex* having white and firm gleba are also edible although it's very much uphill task to obtain the fruiting bodies in this condition (Rinaldi and Tyndalo, 1985). Similarly, the immature puffballs of *Calvatia craniiformis* with a firm, white gleba and mild odor prior to formation of spores are also edible (Meuninck, 2017). The gasteroid *Calvatia gigantea* is also known to have polyunsaturated fatty acids, proteins, carbohydrates, gentisic acid (a phenolic compound) and hexanal (an aroma compound) that make it a capable source of healthy food supplements (Kıvrak *et al.*, 2014).

Dictyophora indusiata (bamboo mushroom) is consumed in China due to its nutritive bioactive compounds that benefits eyes and cardiovascular system and for its moderate medicinal values. The fungus consists around twelve vital metal ions, seven essential amino acids, carotenes, water soluble vitamins (thiamine, riboflavin, nicotinic acid and vitamin C) and minerals like calcium and phosphate (Ker *et al.*, 2011). Likewise, the edible mushroom *Lycoperdon perlatum*, also known as "poor man's sweetbread" (due to their flavor and texture), is considered to be a great source of

food till the gleba is homogeneous and white and become inedible when gleba becomes yellow-tinged (Dickinson and Lucas, 1982). The fungus is well known to contains an unusual and exclusive amino acid lycoperdic acid. The study of Colak *et al.* (2009) revealed that the 100 g dried mass of puffballs contains 42 g carbohydrates, 10.6 g of fats and 44.9 g of proteins together with trace number of micronutrients including copper, iron, manganese and zinc. The predominant fatty acids in fruit bodies are mainly the unsaturated one including palmitic, linoleic, oleic and stearic acid (Nedelcheva *et al.*, 2007).

The fruit bodies of *Mutinus caninus*, *Phallus ravenelli* and *P. impudicus* are known to edible in north America (McIlvaine and Macadam, 1900). At immature egg stage, the crisp and crunchy receptaculum of *P. impudicus* with radishy taste is eaten either raw or as pickle and sausages in France and Germany (Zeitlmayr, 1968; Bon *et al.*, 1987). Similarly, the eggs of gasteroid *Clathrus dbarius* were eaten by the Maori tribes of New Zealand with an inappropriate name "thunder dirt". Additionally, some members of the order *Hymenogasterales* are also being consumed as substitute of truffles (Berkeley, 1867). The fungus *Hymenogaster citrinus*, *Rhizopogon luteolus* and *R. rubescens* are considered edible by mycophagists of England and are being eaten by the local population (Ramsbottom, 1953). The order *Lycoperdales* constitutes the majority of edible species including *Calvatia*, *Lycoperdon* and *Bovista* in north America (McIlvaine and Macadam, 1900).

Fruit bodies of *Phallus indusiatus* particularly in eastern part of Asia is well known for its delicacy and are kept reserved for the special occasions. The mushroom has an attractive shape, tender texture and it remains crisp which make them a decent source of food within the localists. The mushroom in its dehydrated form is sold in the local markets and can be used in variety of ways as they can be rehydrated by soaking and can be used in stir-fry's (cooked in small amount of oil), in chicken soups, stuffed etc. (Newman, 1999; Halpern, 2007). The nutritional composition of *P. indusiatus* at egg stage per 100 g of dry fungus weight contains 1:30 ratio of fats and proteins respectively with 20.9 g dietary fiber and mineral composition as calcium (61.0 mg), copper (1 mg), iron (36.6 mg), magnesium (156 mg), manganese (5.1 mg), potassium (153 mg), sodium (5.1 mg) and zinc (133 mg) (Jonathan *et al.*, 2008).

Phellorinia inquinans is well known for edibility and considered to be a source of high nutraceutical compounds (Gehlot *et al.*, 2016). The sporophore contains around 23 % crude proteins, 9% carbohydrates, 13% fibers and trace amount of lipids along with vital minerals like potassium, phosphorus, calcium, and magnesium (Jandaik, 1977; Doshi and Bohra, 2000). The mushroom also contains a good amount of essential and non-essential amino acids (Sharma *et al.*, 2015). Correspondingly, *Podaxis pistillaris* is utilized in India and Arab countries as a culinary and gourmet sustenance (Singh *et al.*, 2006; Gehlot and Singh, 2016). The fruiting body of *P. pistillaris* contains high values of protein and carbohydrates and at the same time have low fat values

that makes them an important supplement to food. The 100 g of the dry weight comprises 14.5 g protein, 1.97 g fats and 77.79 g carbohydrates. Further it also contains phenols, flavonoids, steroids and β -carotene (Mridu and Atri, 2017) and minerals like Na, P, K, Ca and Mg (Abdalla *et al.*, 2016).

The *Rhizopogon roseolus* is also known to be edible in East-Asia particularly in Japan and the commercial cultivation of the mushroom is also been successfully established in Japan and New Zealand (Yun and Hall, 2004). Chittaragi *et al.* (2014) were tried to evaluate the macro nutrient profiles of *Scleroderma bermudense* and observed that wild mushrooms contain dry matter (9.49), protein (36.32%), ash (4.45-10.29%), lipid (2.79%), fiber (11.44%) and carbohydrate (42.55%) while *Geastrum triplex* contain carbohydrates (40.83%), lipid (2.06 %), protein (27.41%) and fiber (3.77%). Abdalla *et al.* (2016) studied proximate composition of *Calvatia cyathiformis* and estimated 40.47% crude protein 3.05% crude fat, 24.83% carbohydrate, 16.25% crude fiber with mineral constituents of sodium, potassium, phosphorus, calcium and magnesium. Agrahar-Murugkar and Subbulakshmi (2005) estimated 27.3% protein, 1.0% lipid/fat and 22.0% fiber with mineral nutrients (Ca, P, Fe, Mn, Cu, Zn, Na, K, Mg and Se) in *Calvatia gigantea*. Pavithra *et al.* (2018) attributed nutritional quality of edible gasteroid wild fungus *Astraeus hygrometricus* and quantified crude protein 16.80%, total lipids 3.55%, crude fibre 14.58% and total carbohydrates 46.17% with adequate quantity of some important minerals and six essential amino acids. Therefore, the mushroom is a low energy healthy food that can be utilized as a protein supplements which have adequate amount of carbohydrates, essential amino acids, fibers, important vitamins and minerals.

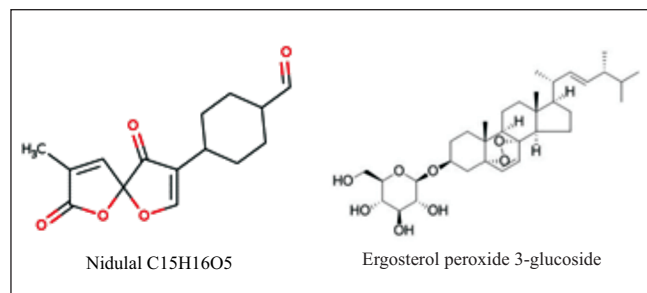
PHARMACEUTICAL COMPOUNDS/ THERAPEUTIC SIGNIFICANCE

Anti-cancerous and anti-tumor compounds

The extract of *Astraeus hygrometricus* with a chief polysaccharide AE2 was reported to have profound inhibitory ability against several tumor line *in-vivo* (Mallick *et al.*, 2010). The AE2 is composed of mannose, glucose and fucose in a 1:2:1 ratio and it is known to be activated by a signal transduction pathway mediated by mitogen-activated protein kinase pathway (Mallick *et al.*, 2011). Additionally, the extract was also reported to stimulate the growth of splenocytes, thymocytes, and bone marrow cells in mice (Mallick *et al.*, 2011). The puffballs of *Calvatia* are the chief source of calvacin (an anti-tumor mucoprotein), although found in very small quantities but the compound is known to stimulate the immune cells (Coetze and Wyk, 2009).

Mannan having the molecular weight of 620 kDa have been extracted from the liquid cultures of *Dictyophora indusiata* exhibited strong anticancer bioactivity (Hara *et al.*, 1982). Similarly, the polysaccharides from fruiting body showed immuno-suppressive effect on prostate-cancer-associated fibroblasts (Habtariam, 2019). Erkel *et al.* (1996) reported two vital cytotoxic sesquiterpene compounds nidulal and niduloic acid from *Nidula candida* having significant role in

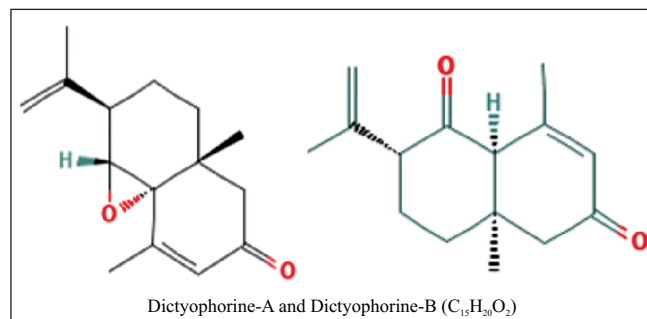
the upregulation of human promyelocytic leukemia cells differentiation. Moreover, fungus is also known as a potent scavenger of superoxide anions that shields the lipoproteins of plasma membrane from peroxidation predicting its anti-cancerous and immuno-stimulatory effects (Wei, 2005).



Gasteroid mushroom *Phallus indusiatus* is known to have a bioactive polysaccharide that have a structure constituted chiefly with D-mannopyranosyl residues linked with α -1 \rightarrow 3 linkages together with traces of 1 \rightarrow 6 linked D-mannopyranosyl residues (Ukai *et al.*, 1980). The polysaccharide was found to have a potent tumour-suppressing ability against subcutaneously implanted sarcoma (Ukai *et al.*, 1983). Kovacs *et al.*, (2018) has isolated and characterized an antiproliferative secondary metabolites from the potato earthball mushroom, *Scleroderma bovista*. He observed that the lanostane-type derivatives and ergosterol peroxide 3-glucoside exerted significant antiproliferative property on one or more human cancer cell lines on MTT assay.

Immuno-modulatory properties

The extract of *Astraeus hygrometricus* was found to encourage cell of immune system in mouse where it stimulated the activity of natural killer cells, macrophages to produce nitric oxide and enhanced production of cytokines (Chakraborty *et al.*, 2004). Under in-vitro conditions the polysaccharides extracted from the fruiting bodies of *Dictyophora indusiata* showed a significant immuno-stimulatory effect as these were found to induced a proliferative response whereas, the cytokines, No synthase and nuclear factor k-light-chain-enhancer of B-cells as the markers of macrophages were also upregulated (Deng *et al.*, 2016). In same context the study of Han *et al.*, (2017) on prostate cancer fibroblasts showed that the fungal polysaccharides not only induced the proliferation of lymphocytes but also upgraded the growth of suppressed CD4+/CD8+ T cells.



Dictyophorine-A and Dictyophorine-B are the two novel sesquiterpenes that have been isolated from the fruit bodies of fungus *Phallus indusiatus* (Kawagishi *et al.*, 1997). The compounds are characterized as the pioneer eudesmane derivatives with potential to induce the nerve-growth-factors (NGF) in astrocytes, the cells which forms the blood brain barriers (Kawagishi *et al.*, 1997). Additionally, Liu (2005) reported the quinazoline derivatives (Dictyoquinazol A, B, and C) isolated from the fungus have a shielding effect on neurotoxins exposed mouse neurons.

Anti-inflammatory activity

Inflammation in general is defined as a complex biological process, induced by natural microbial infection or by accidental damage of tissues (García-Lafuente *et al.*, 2009). The inflammation may leads to various rheumatic disorders such as fever, arthritis, ankylosing spondylitis (Anilkumar, 2010). In other word the Inflammation can be defined as body defense activities in order to limit or eliminate the spread of injurious agent; oedema, leukocyte infiltration, and granuloma formation etc. can be observes as a byproduct of the process (Amdekar *et al.*, 2012). There are many of the drugs which available to counter the inflammatory responses but these are toxic too that have a severe side effect including various gastrointestinal problems (Bisht *et al.*, 2014). Therefore, there is need of natural anti-inflammatory drug with significant functionality and without imparting any side effects.

The ethanolic extract of *Astraeus hygrometricus* fruit bodies showed anti-inflammatory activity comparable to the drug diclofenac in laboratory experiments (Biswas *et al.*, 2010). Furthermore, the two water soluble glucans, having β -1 \rightarrow 6 branches linked to β -(1 \rightarrow 3)-D glucose main frames, isolated from the liquid cultures of *Dictyophora indusiata* showed significant anti-inflammatory responses (Hara *et al.*, 1982).

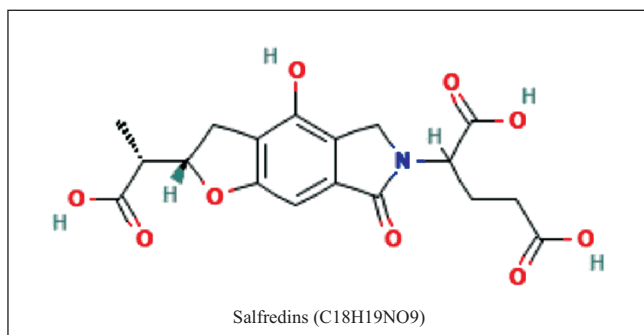
A β -glucan protein complex isolated and characterized from *Geastrum saccatum* found to inhibit the inflammatory enzymes like nitric oxide synthase and cyclooxygenase and thus proved to be a substantial anti-inflammation agent (Dore *et al.*, 2007). The lipoxygenase (LOX) enzymes are responsible for the development of leukotrienes (LTs) that increases under rheumatic conditions and plays a vital role in pathophysiology of various inflammatory diseases. The inhibition of LOX pathway directly prevents the formation of LTs (Yahaya and Don, 2012). The methanolic extract from *Geastrum fimbriatum* has been tested for lipoxygenase inhibition essay and observed to have 12.92% inhibition (Sarac *et al.*, 2019). Similar, reports were also obtained from the study of Dore *et al.* (2007) on *Geastrum saccatum* extract that found to be a potential inhibitor of enzyme cyclooxygenase.

The puffballs of *Mycenastrum corium* are although very thick due to their outer periderm but these puffballs are also used for their alleged anti-inflammatory properties in Mexico (Pérez-Silva *et al.*, 2015). A bioactive polysaccharide β -D-glucan, named as T-5-N isolated in alkaline extracts from the fruit bodies of *Phallus indusiatus* contains significant anti-

inflammatory properties (Ukai *et al.*, 1983).

Anti-diabetic activity

The species of genus *Bovista* are suggested to improve diabetes mellitus (Jain, 1997). The whole fruiting body extract of *Calvatia gigantea* showed a significant in-vitro alpha amylase inhibitory activities and *in-vivo* antidiabetic activity that underlined the facts that the gasteroid fungi could be used potentially as a functional antidiabetic (Ogbole *et al.*, 2019). Matsumoto *et al.* (1995) extracted new bioactive compounds designated as salfredins from the liquid cultures of the fungus *Crucibulum laeve*. salfredins have structural resemblance with benzofuran and chromene containing cyclic amide or lactone ring structures. These salfredins have therapeutic use as it inhibits the enzyme aldose reductases responsible for the formation cataracts during the advanced stages of diabetes mellitus (Kyselova *et al.*, 2004; Srivastava *et al.*, 2005).



The alkali extracted polysaccharides from *Dictyophora indusiata* was proved to have a direct glucose lowering effect and also regularized the serum adiponectin level, insulin and leptin (Wang *et al.*, 2019).

Antioxidant properties

The reactive oxygen species in form of free radicals including superoxides, hydroxyl ions are produced as a byproduct of human physiological processes that causes oxidative stress by reacting with different biomolecules such as proteins, lipids, DNA etc. (Gülçin *et al.*, 2002). Although the living organism itself possesses variety of endogenous defense system to counter the oxidative damages but the supportive exogenous supply of antioxidants is always essential that scavenge the free radicals and maintains the normal physiology of the cells (Halliwell and Gutteridge, 2015). Not only in the living systems but also the preservative foods require these antioxidants to counter the peroxidation and therefore synthetic antioxidants like propylgallate, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT) and tertabutylhydroquinone are used as preservative. These artificial scavengers cause variety of damages such as liver-cirrhosis and carcinogenesis and affects the normal physiology (Hettiarachchy *et al.*, 1996). Therefore, there is need of antioxidants from natural compounds without any adverse effects and none is more important than mushrooms that produce variety of bioactive compounds with a known and potential antioxidant properties (Caleja *et al.*, 2017).

The ethanolic extract of *Astraeus hygrometricus* fruit body have shown significant antioxidant activity. Clinical studies have revealed the hepatoprotective properties in mouse as the extract found to restore the diminished levels of superoxide dismutase and catalase (antioxidant enzymes) in mouse exposed with carbon tetrachloride (Biswas *et al.*, 2011). Likewise, the polyketide-antioxidants such as cyathusals, pulvinatal and cyathuscavin was identified in the liquid cultures of *Cyathus stercoreus* which showed antioxidant and nucleic acid protection activities (Kang *et al.*, 2007). Polysaccharides from *Dictyophora indusiata* showed a potent antioxidant activity in *Caenorhabditis elegans* under the oxidative stress conditions induced with parquat where it decreased the level of reactive oxygen species (ROS) and malondialdehyde (MDA) and showed increased levels of SOD (Zhang *et al.*, 2016).

Sevindik *et al.* (2017) determined the total antioxidant status (TAS), total oxidant status (TOS), oxidative stress index (OSI) of *Geastrum pectinatum* mushroom and found that the methanolic extract of the mushroom showed a significant TAS value and could be consumed as an alternative antioxidant source. In the same context, chemical constituents and antioxidant activity of the Arctic mushroom *Lycoperdon molle* has been exploited in the study of Singh *et al.* (2012) where, in order to determine the in-vitro antioxidant potential, fungal fruiting bodies were extracted in 20 ml of methanol, ethanol, acetone and DMSO using a soxhlet extractor. The extracts were tested for free-radical scavenging (FRS) activity, inhibition of lipid peroxidation (ILP) and Trolox equivalent antioxidant capacity (TEAC) and concluded to have more than 50% potential for FRS, ILP and TEAC with maximum FRS in methanolic extract (89%). The strong correlation between the antioxidative potential and higher polysaccharide concentration has also been established that also corroborate the findings of Liu *et al.* (1997), who also reported that the polysaccharides have the ability to scavenge free radicals.

The extracts of the fruit body of *P. indusiatus* contains antioxidant properties due to the presence of polyphenols, known to counter the cellular damage due to the formation of ROS (Mau *et al.*, 2002). Similarly, the extracts of mushroom *Phellorinia herculeana* was analyzed for radical scavenging effect by the DPPH assay. The extracts showed inhibition values greater than 90% (concentration of 500 mg) comparable with the activity of the reference compound ascorbic acid (Al-Fatimi *et al.*, 2005). Furthermore, the gasteroid mushroom *Podaxis pistillaris* is also considered as an important antioxidant source due to presence of secondary metabolites such as phenols, flavonoids, steroids, β -carotene and lycopene in its fruit bodies (Mao, 2000; Mridu and Atri, 2017).

Antibiotics compounds

The macro-fungi are also rich sources of natural antibiotics. The glucans as a component of their cell wall along with several secondary metabolites showed antibacterial, antifungal and anti-viral activities and are known to be

potential immuno-modulators (Cohen *et al.*, 2002). Although the medicinal values of these wild mushrooms have been exploited nowadays but they have a history of traditional medicine for curing various types of diseases (Iwalokun *et al.*, 2007).

Gasteroid *Cyathus striatus* in 1971 was first time reported to have ability for the production of compounds with an indole ring collectively known as indolic-substances in addition to a complex of diterpenoid antibiotic compounds known as cyathins (Allbutt *et al.*, 1971). These indolic substance were later on named as striatins with potential antibiotic activity against imperfect fungi, gram positive and negative bacteria (Anke *et al.*, 1977).

Akpi *et al.* (2017) studied the antimicrobial activity and phytochemical characteristics of the crude extract of *Lycoperdon perlatum*. The phytochemicals analysis using the ethanol, methanol and water extracts of puffballs suggested the presence of diverse bioactive compound including flavonoids, saponins, protein, carbohydrate and glycosides. The agar gel diffusion methods were applied in order to evaluate the antimicrobial activity. The aqueous extract showed retardation in growth of all the tested microbes including *Staphylococcus aureus*, *Bacillus cereus*, *Candida albicans* except *Pseudomonas aeruginosa* while the methanol and ethanol extracts was found to be most effective as they inhibited all the tested organisms. The outcome suggests the gasteroid as a broad-spectrum antimicrobial agent. Moderate antibiotic activity were also reported from *Nidula candida* due to two bioactive fifteen-carbon sesquiterpene compounds, nidulal and niduloic acid (Erkel *et al.*, 1996).

Addition of fungal extract of *Phallus indusiatus* in the liquid broths prevent it from spoiling for many days. The extracts were also tested against numerous human pathogens and reported to be a decent substitute for the antibiotic's ampicillin, tetracycline, and nystatin (Habtemariam, 2019). An antibiotic albaflavene was later on isolated from the fungus that's chemically a sesquiterpene already known from *Streptomyces albidoflavus* (Huang *et al.*, 2011).

Antibacterial activity

An increase in the resistance to existing antibiotics put an enormous load on pan-scientific community for innovation and discoveries of the new emerging antibiotics and for this none have more potential than the herbs, plants and mushrooms (Iftikhar *et al.*, 2011). The production of antimicrobial metabolites from the mycelial or spore culture of wild mushroom has been reported substantially throughout the world (Hasan *et al.*, 2015). *Cyathus helenae* has been shown to produce a complex of metabolites so called cyathin complex that has antimicrobial properties and showed to inhibit the growth of actinomycetes, bacteria and some fungi (Allbutt *et al.*, 1971; Ayer and Taube, 1972). In 2008, five lanostane-type triterpenes were isolated and identified from the fruit bodies of *Astraeus pteridis*; two of these compounds had an inhibitory effect on the growth of the pathogen *Mycobacterium tuberculosis* with antituberculosis activity (Stanikunaite *et al.*, 2008).

The liquid extract of *Dictyophora indusiata* exhibited a weaker activity against microzymes and molds but found to have immensely potential anti-bacterial activity (Hao *et al.*, 2008). The well-known antibiotic albaflavene has also been isolated from *D. indusiata* which is a module of streptomycetes and its biosynthesis pathway and activities have been the subject of many microbial studies. Chittaragi *et al.* (2013) demonstrated the antibacterial activities of petroleum ether, chloroform and methanol extract of mushroom *Geastrum triplex* using agar well diffusion method on three plants and six human pathogenic bacteria. They observed maximum antibacterial activity of chloroform extracts against *Xanthomonas campestris* and of methanol extract against *Agrobacterium tumefaciens*. Further concluded that chloroform extracts showed promising antibacterial activity against plant pathogens then other two whereas, the petroleum ether extract were found more effective against human pathogenic bacteria.

Methanolic extracts of *Lycoperdon umbrinum* with terpenoids as key component has showed a significant anti-bacterial activity against numerous human pathogens including *Bacillus subtilis*, *Escherichia coli*, *Salmonella typhimurium*, *Staphylococcus aureus*, *Streptococcus pyogenes*, and *Mycobacterium smegmatis* (Dulger, 2005) and found moderately effective against *Enterococcus faecium* and *Staphylococcus aureus* (Suay *et al.*, 2000). Similarly, the extract from species *L. perlatum* showed antibacterial activity analogous to the antibiotic ampicillin and inhibits the growth of numerous human pathogens including *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa* (Ramesh and Pattar, 2010). Additionally, the study of Dulger (2005) found similar antagonistic activity against *Mycobacterium smegmatis*, *Salmonella enterica* and *Streptococcus pyogenes*.

The extracts from the fruit bodies of fungus *Mutinus elegans* showed an antibacterial activity against the five human pathogenic bacteria *Salmonella typhimurium*, *Bacillus cereus*, *B. subtilis*, *Escherichia coli*, and *Staphylococcus aureus* (Coletto and Lelli, 1998). Likewise, dichloromethane, methanol and water extracts of mushroom *Phellorinia herculeana* showed a potential antagonistic activity against *Streptomyces aureus* and *Bacillus subtilis* in agar diffusion assay whereas, the bacteria *Mariniluteicoccus flavus* and *Candida maltosa* found to be resistant against all three extracts. Additionally, the dichloromethane and methanol extract of fungus found to be effective against *E. coli* and *P. aeruginosa* but both showed resistance against water extracts (Al-Fatimi *et al.*, 2005). In the same context, Gehlot *et al.* (2018) tested the ethyl acetate extract of *P. herculeana* fruiting bodies for their efficacy against five human pathogenic bacteria *E. coli*, *Vibrio cholera*, *Salmonella enteric typhimurium*, *S. aureus* and *Enterococcus faecalis* using the agar gel diffusion method and observed that crude extract checked the growth of all the five pathogenic bacteria, further, gas chromatography mass spectroscopy analysis of crude suggested the presence of decane, 1-chloro- and dodecane, 1-chloro responsible for this activity.

Antagonistic activity of *Podaxis* sp. was determined against

the number of pathogenic gram negative and positive bacteria like *P. aeruginosa*, *Proteus mirabilis* (Panwar and Purohit, 2002), *S. aureus*, *Micrococcus flavus*, *B. subtilis*, *Proteus mirabilis*, *Serratia marcescens* and *E. coli* (Al-Fatimi *et al.*, 2006). Al-Fatimi *et al.* (2006) identified three possible antibacterial compounds as epidi-thio-diketo-piperazines viz., epicorazine A, epicorazine B and epicorazine C.

Antifungal activity

A number of *Cyathus* species are reported to be effective against some of the human pathogens like *Cryptococcus neoformans* (infects lungs and central nervous system), *Aspergillus fumigatus* (causes chronic pulmonary infections) and *Candida albicans* (causes genital yeast infections) (Liu and Zhang, 2004). Liquid broth cultures of *Geastrum* species were found to have effective anti-microbial activities against pathogenic fungi *Candida krusie*, *C. albicans*, and *Trichophyton mentagrophytes* *in-vitro* in agar cup diffusion method on potato dextrose broth medium (Panda *et al.*, 2017).

A number of fungal species including *C. albicans*, *Aspergillus fumigatus*, *Botrytis cinerea* and *Verticillium dahliae* showed a significant retardation in their growth when subjected to the puffballs extract from the mushroom *Lycoperdon perlatum* (Pujol *et al.*, 1990). The extract from *Mutinus elegans* showed a potential anti-fungal activity against the yeast *C. albicans* (Coletto, 2006). In the same context, the ectomycorrhizal fungi *Scleroderma citrinum*, *Lycoperdon perlatum* and *Pisolithus albus* was found to have antifungal potential against *Alternaria solani*, *Botrytis* sp., *Fusarium oxysporum*, *Phytophthora* sp., *Pythium* sp. and *Rhizoctonia solani* in various capacity in *in-vitro* conditions (Mohan *et al.*, 2015). These species were further quantitatively assessed for the production of enzyme chitinase and found to have around 38.99 µg/ml in *S. citrinum* and 32.38 µg/ml of chitinase in *P. albus* (Mohan *et al.*, 2015). Similarly, Soytong *et al.* (2014) isolated two bioactive lanostane-type steroid compounds named 4,4-dimethoxy-methyl vulpinate (DMV) and 4,4-dimethoxyvulpinic acid (DMVA) from fruiting body of *S. citrinum* which were found significantly inhibiting the growth of *Phytophthora palmivora* and *Colletotrichum gloeosporioides*.

Two important antibiotic compounds named pisolithin-A (p-hydroxy benzoyl formic acid 2-(4'-hydroxyphenyl) 2-oxoethanoic acid) and pisolithin-B (p-hydroxymandelic acid (R)-(-)-2-(4'-hydroxyphenyl)-2-hydroxyethanoic acid) has been extracted from the cultures of *Pisolithus tinctorius*. These compounds are well characterized for their inhibitory potential towards the germination of spores of many plant pathogenic fungal species like *Phytophthora* sp., *Rhizoctonia solani*, *Fusarium solani*, *Pythium ultimum* (Tsantrizos *et al.*, 1991). These gasteroid fungi not only act as biocontrol agent against phytopathogens but also can contribute in plant growth promotion by production of various phytohormones including indole acetic acid, cytokinin and gibberellin (Ek *et al.*, 1983).

Other therapeutic uses

The cyathane-diterpenoids isolated from the *Cyathus* spp. has

been reported to stimulates nerve growth factor (NGR) synthesis that ensures their therapeutic potential for treatment of Alzheimer's disease (Krzyczkowski *et al.*, 2008). The patients with recurring breast cancer have tendency to develop the blood clots in vein (venous thrombosis) that contributes majority of the death incidence in cancer survivors, therefore, they have to kept lifelong on the anticoagulants. The extract from the fruit bodies of fungus *Phallus impudicus* have potential to reduce the clumping of platelets and thus can be utilize as supportive preventive nutrition (Kuznecov *et al.*, 2007).

Fruit bodies of *Scleroderma polyrhizum* comprise steroids ergosta-4,6,8(14) 22-tetraen-3-one and 5 α ,8 α -epidoxysterosta-6,22-dien-3 β -ol along with unsaturated fatty acids palmitic and oleic acids and are used as a conventional Chinese medicine for the treatment of detumescence and hemostasis (Xianling *et al.*, 2005).

The spore mass of *Podaxis pistillaris* and *Geastrum triplex* are used in skin disease and skin burns (Kumar *et al.*, 2017). The spore mass of *Astraeus hygrometricus* mixed with mustard seed oil is used as an ointment for burn cases (Rai *et al.*, 1993). The spores of *Pisolithus arrhizus* are used as medicine for treatment of wounds (Manna *et al.*, 2014). *Calvatia gigantea* is used in stomach upset and to cure stomach pains in women during menstruation (Thangaraj *et al.*, 2017). The spore of *Lycoperdon perlatum* is used to dress wounds for quick healing of wound and also to stop bleeding (Pala *et al.*, 2013). *Bovista plumbea* is used for the treatment of sores, ulcers and skin infection (Devi *et al.*, 2016). *Rhizopogon villosulus* is used against kidney stone, urinary tract infections and also to cure fatty liver and asthma (Malik *et al.*, 2017).

Gasteroid fungi are not only responsible for the production of substance that humans can use directly as medicines but also these are the versatile tools in the vast field of medical research. The utilization of *Bovista stevenii* spores as substance responsible for promoting the process of healing through the re-formation of scar tissues (Scarpa, 2004). Similarly, *Mycenastrum corium* can be used remedially in Mexico as a hemostatic and also as a tonic for throat and lungs (Pérez-Silva *et al.*, 2015).

The gasteroid fungi are also known to have products which were previously not known from the biological origins. Three exclusive triterpene derivatives of 3-hydroxy-lanostane having δ -lactone (a six-membered ring) in the side chain have been isolated from *Astraeus hygrometricus* fruit bodies that represents a novel chemical feature which is not previously known from the mushrooms of *Basidiomycotina*. In addition to this, another different steryl ester (3 β , 5 α -dihydroxy-(22E, 24R)-ergosta-7,22-dien-6 α -yl palmitate) also known to be present in the liquid cultures of gasteroid (Hong-Jun *et al.*, 2007). The *Bovista* has proven to be a significant curative in head affection characterized by sensation as if the head were enormously increased in size. The spores of *Bovista* is also known to restrict blood circulation through the capillaries and found accompanying with menstrual irregularity or trauma. It also known for causing suffocation and might be beneficial in remedying

asphyxiation resulting from inhalation of charcoal fumes (Farrington, 1999). Other benefits of *Bovista* includes improvement of awkwardness in speech and action, stuttering or stammering in children, palpitation after a meal, diabetes mellitus, ovarian cysts, and acne due to cosmetics (Jain, 1997).

Kakumyan *et al.* (2020) collected the volatile organic compound emitted from different stage of fruit bodies of the stinkhorn fungus *Pseudocolus fusiformis* and analyzed using the gas chromatography-mass spectrometry (GC-MS). They observed 3-methyl-butanol, 4-methyl-phenol, and dimethyl tetra-sulfide in addition to dimethyl disulfide, trisulfide and tetra-sulfide as the major organic volatiles in mature fruit bodies. *Cavatia gigantean* and *C. caelata* possess, when burnt, anaesthetic properties similar to those of chloroform (Berkeley, 1867).

APHRODISIAC PROPERTIES

The two *Cyathus* species named *C. limbatus* and *C. microsporus* are used as a source of vitality (aphrodisiac) by the local tribes of Colombia and Guadeloupe respectively. Although the actual effect of these fungus on physiology is not clearly known (Brodie, 1975). Likewise, fruit bodies of *Phallus impudicus* is fed to the young bulls in Northern Montenegro due to its potent aphrodisiac property whereas, it is gently rubbed on the necks of bulls prior to their combat in order to make them strong (Davis *et al.*, 2012). Similarly, The unopened stinkhorns, known as "ghost's or daemon's eggs" among local population of Thuringia, is used as dried powder in drinks as an aphrodisiac (Lightfoot, 1789). In the same context, Holliday and Soule (2001) claimed that some species of *Phallus* found in Hawaii is aphrodisiac to women and just inhaling the mushroom can cause spontaneous orgasm. Although, the study never reproduced and proved by any other study.

DYE/ PIGMENT COMPOUNDS

Red colour of *Calostoma cinnabarinum* is ascribed to the presence of pigment 'calostomal' with IUPAC nomenclature of all-trans-16-oxohexadeca-2,4,6,8,10,12,14-heptaenoic acid (Gruber and Steglich, 2007). Similarly, the fruit bodies of *Lycoperdon perlatum* contain the pigment melanin (Almendros *et al.*, 1987).

The major pigments responsible for the conversion of fruits from orange to red are carotenes, predominantly lycopene and beta-carotene. The lycopene is identified as a chief pigment in fungus *Clathrus archeri*, while beta-carotene is the predominant pigment in the family *Phallaceae* (Fiasson and Petersen 1973). Aulingeret *et al.*, (2001) isolated a poly-unsaturated organic compounds (polyene pigment) named melanocrocine from the fruit body and mycelial cultures of subterranean fungus *Melanogaster broomeianus*. Melanocrocine is a N-acyl derivative of L-phenylalanine methylester.

Genus *Scleroderma* is mainly consisting of gasteroid ectomycorrhizal (ECM) species having spores which are characteristically reticulate to echinulate globose and named as earthballs (Sims *et al.*, 1995). The alkaline extraction

method was applied for the isolation of melanin pigments from the moist gleba of *S. citrinum* fruiting bodies further refined by acid hydrolysis and washing with organic solvents. The pigment also showed ROS scavenging and antimicrobial activity (Łopusiewicz *et al.*, 2018). Melanin is generally known as brown and black pigments derived from oxidation of monophenols (Solano, 2014). Therefore, melanin's from *S. citrinum* possess notable therapeutic action and could be applied in the food, cosmetics and pharmaceutical industries. The pigment found in the fruiting body of *S. citrinum* are sclerocitrin, norbadione A, xerocomic acid and badione A (Winner *et al.*, 2004).

Furthermore, gasteroid fungus *Pisolithus arrhizus* is also being used as a natural dye for coloring of clothes (Roberts and Evans, 2011). *P. tinctorius* is locally referred as 'Dye Ball' due to the presence of purple and yellow dyes in the fruiting bodies and is utilized extensively for dyeing cotton and wool, in the Canary Islands and parts of Italy (Rolfe and Rolfe, 1925). Tan *et al.* (1978) first reported the production of dark-colored pigments, phenolic, p-hydroxybenzoic and humic acid from the fruit bodies of *P. tinctorius* and similarly the study of Gill and lally (1985) firstly evaluated the presence of pigment norbadione-A from the sporophores of fungus accountable for the intense pigmentation of fruit bodies.

AGRICULTURAL PRODUCTS

Cyathus stercoreus has been reported to have ligninolytic and cellulolytic activities which selectively degrade lignin in agricultural byproducts such as wheat straws, leaving great part of cellulose intact. It results in increase in digestible carbohydrates for ruminants which improves the quality of food source (Wicklow *et al.*, 1980; Akin *et al.*, 1995).

The extract of *Podaxis pistillaris* harbors a significant fibrolitic activity and thus make them a probiotic, a food supplement having living bacteria (Vasquez-Davila, 2017). The exogenous fibrolytic enzymes are the bio-catalyst used for the eradication of non-nutritive products from the foods of monogastric and ruminant species (Bedford, 2000). Squeezing the spore sac of *Geastrum saccatum* and *G. triplex* released spores into bee hives which anesthetize bees for about 30 min facilitating harvest of honey (Tibuhwa, 2012).

INDUSTRIAL PRODUCTS

The enzymes such as laccase and manganese peroxidase extracted from the liquid cultures of *Cyathus stercoreus* are well known to have potential applications in pulp and paper industries. The ligninolytic potential of these enzymes are used for removal of lignin from paper pulp (Sethuraman *et al.*, 1999). In addition to this, the species *C. bulleri* contains three enzymes (lignin peroxidase, manganese peroxidase, and laccase) that not only contributes in the paper and pulp industries but also have abilities to enhance the digestibility and protein contents in forage (Vasdev *et al.*, 1995). Further the enzyme laccase also has potential of breaking down the phenolics that might be useful in excavating toxic materials from the effluents of dyes and textile industries thus making it a good environmentally friendly practice (Chhabra *et al.*, 2008). The liquid cultures of *C. stercoreus* and *C. pallidus*

was reported to have biodegradable activity to degrade explosive 2,4,6-trinitrotoluene (TNT) and RDX (hexahydro-1,3,5-trinitro-1,3,5-triazine) by Bayman *et al.* (1995).

The commercial pesticides manufacturers utilize the bioactive compounds from the species of the genus *Nidula*. These compounds in their acetate form include raspberry ketone with chemical name 4-(p-hydroxyphenyl)-2-butanone as a major component of raspberry flavor and insect attractor. It also shows structural resemblance with sex pheromones of female melon fly that attracts male insects (Ayer and Singer, 1980). Furthermore, fruit bodies of mushroom *Lycoperdon perlatum* contains 14 mg/kg cinnamic acid of fungal dry weight (Barros *et al.*, 2009) that can be utilized in various industries as flavorings, synthetic indigo, perfume industries and also as a precursor of sweetener aspartame (Garbe, 2000).

Hydroxy methyl furfural is a chemical, extracted from the fruit bodies of fungus *Phallus indusiatus*, having the tyrosinase inhibitor activity (Sharma *et al.*, 2004). The enzyme responsible for catalyzing the pioneer step in the process of melanogenesis particularly in mammals and unwanted browning reactions in harvested fruits that make this inhibitor a vital component for cosmetic and food industries (Chang, 2009). The starches from the fruiting bodies of fungus with various physicochemical and functional properties plays a vital role in food and other industries. These physiochemical properties can be influenced by the ratio of amylose and amylopectin. These contribute as thickeners in food processing and also have applications in cosmetics, paper coatings, laundry and biofilms (Kong *et al.*, 2009). *Scleroderma citrinum* is known to grow vigorously on dead and decaying woods as they are lignocellulolytic fungi. The total starch content from the fruit bodies was evaluated to have high amylose and comparatively low amylopectin. The gelatinization temperature was found to be in optimum range and the sample showed stable dextrin production which suggests the potential applicability of these starches in non-food industries (Aniekemabasi *et al.*, 2018).

ETHNOMYCOLOGICAL KNOWLEDGE OF ETHNIC TRIBE

Astraeus hygrometricus is traditionally used as a hemostatic agent in China. The powdered dust prepared from spore is applied directly on cuts and wounds to stop bleeding (Hobbs, 1995). The tribes Biga and Bharia in the forests of Madhya Pradesh (India) are also very much acquainted to medicinal properties of the fungus. The blend of spores in mustard oil is used by them against the burns (Rai *et al.*, 1993). Similarly, decoction (boiled extract) of fungus *Cyathus stercoreus* is used as a Chinese medicine to reduce the symptoms of gastralgia (Liu and Bau, 1980).

Indigenous people of the South-Eastern woodlands of the United States utilize the fruiting body of *Geastrum triplex* as prophylactic on the umbilici of newborns after birth until the fell off of umbilical cord. Further, it is also used to reduce inflammation and swelling of respiratory tracts (Ying, 1987). Likewise, stinkhorns of *Phallus indusiatus*, due to their

unique shapes, are consumed during the divinatory ceremonies in Mexico (Læssøe and Spooner, 1994). The mushroom is designated as Akufodewa by the Yo-ruba tribes of Nigeria which means “die-for-hunter-search” because the unusual odor of fruit bodies attracts hunter who mistakes them for dead animals (Oso, 1975). From the Chinese Tang Dynasty, the fungus is utilized for the treatment of inflammations and neural diseases as mentioned in pharmacopoeia. Similarly, it has been used by Miao people in China for the treatment of various conditions including cough, dysentery, leukemia, laryngitis and leucorrhea, oliguria, diarrhea, hypertension and hyperlipidemia (Ker *et al.*, 2011).

The mushroom *Phellorinia herculeana* is known to have numerous medicinal properties and exploited throughout the globe for their health benefits (Gehlot *et al.*, 2018). The basidiospores upon their maturity are traditionally applied on the fresh wounds, on cracked bone and also contributes in management of diabetes and cardiac conditions (Sharma and Doshi, 1996; Gehlot and Gehlot, 2020). The traditional pharmacological literature of many countries comprises the name of *Podaxis pistillaris* (Muhsin *et al.*, 2012) with numerous health benefits such as treatment of skin disease, wound healing, sunburn, inflammation and their antibacterial effects (Diallo *et al.*, 2002; Al-Fatimi *et al.*, 2006). Therefore, both *P. herculeana* and *P. pistillaris* are being sold out in market by ethnic communities with high price (Gehlot, 2016).

Four gasteroid fungi (*Lycoperdon echinatum*, *Bovista nigrescens*, *Pisolithus arrhizus* and *Scleroderma citrinum*) are known traditionally for the treatment of skin diseases and wound healing (Chang and Miles, 2004). The fumes of burning fruit bodies of puffball *Calvatia gigantea* was believed to have tranquilizing properties and used by the tribal people to calm the bees to facilitate the collection of the honey (Dickinson and Lucas, 1979). This sedative property is not due to the presence of any anesthetic compound rather due to an excess of carbon dioxide. According to Rolfe and Rolfe (1925), the hygroscopic nature of mushroom *Astraeus hygrometricus* leads to the use of the species as a hygrometer. Similarly, in some parts of China, the boiled extract of *Phallus* and *Dictyophora* are used as a preservative to preserve the food for shorter durations (Ying, 1987).

CONCLUSION

Although the much literature available on the bioactive compound of mushrooms but the information about the gasteroid bioactives is either not available or if available it is in a very discreet form therefore, the present review aims to compile a concise report on bioactive compounds of gasteroid fungi. In this study not only the nutritive but also pharmaceutical activities of gasteroid fungi have been explored along with their industrial and ethnomycological applications. The bioactive compounds extracted from the fruiting bodies of these fungi are potential anticancerous, antifungal, antimicrobial, antioxidant and antifungal agents. As far as the industrial applications are concern a number of dyes, ligninolytic and cellulolytic compounds, exchangers of

toxic materials from the effluents of dyes and textile industries have also known to be extracted from these fungi.

FUTURE PROSPECTS

The macrofungi (mushrooms) are well known for the production of wide range of natural products as a part of their secondary metabolism like polyketides, glucans, peptides terpenes/terpenoids etc. The interest in these bioactive compounds has been considerably increased nowadays, as most of these fungal metabolites have their medicinal, industrial and agricultural applications. Although plenty of synthetic drugs are available today in the form of antibiotics, antimycotics, nematocidal, etc. but at same time their continuous application also increasing resistance of microorganisms towards these drugs. Therefore, there is need for the exploration of new antimicrobial drugs from the natural resources. In this review, we aim to provide a report on the potential of gasteroid fungi in the food, pharmaceuticals, agricultural industries which otherwise unavailable for the researchers. The study will help to understand the significance of the gasteroid macrofungi and will call the attention of researcher to further explored bioactive compounds from these gasteroid.

REFERENCES

- Abdalla, R.R., Ahmed, A.I., Abdalla, A.I., Abdelmaboud, O.A.A., Khiery, N.T.M.A., Elriah, N. and Saeed, M. 2016. Some wild edible and medicinal mushroom species at Khartoum and Sinnar states-Sudan. *J. Microb. Biochem. Technol.* **08**: 503-506.
- Agrahar-Murugkar, D. and Subbulakshmi, G. 2005. Nutritional value of edible wild mushrooms collected from the Khasi hills of Meghalaya. *Food Chem.* **89**: 599-603.
- Akgul, H., Sevindik, M., Coban, C., Alli, H. and Selamoglu, Z. 2017. New approaches in traditional and complementary alternative medicine practices: *Auricularia auricula* and *Trametes versicolor*. *J. Tradit. Med. Clin. Naturop.* **6**: 239.
- Akin, D.E., Rigsby, L.L., Sethuraman, A., Morrison, W.H., Gamble, G.R. and Eriksson, K.E. 1995. Alterations in structure, chemistry, and biodegradability of grass lignocellulose treated with the white rot fungi *Ceriporiopsis subvermispora* and *Cyathus stercoreus*. *Appl. Environ. Microbiol.* **61**: 15-91.
- Akpi, U.K., Odoh, C.K., Ideh, E.E. and Adobu, U.S. 2017. Antimicrobial activity of *Lycoperdon perlatum* whole fruit body on common pathogenic bacteria and fungi. *Afr. J. Clin. Exp. Microbiol.* **18**: 79-85.
- Al-Fatimi, M., Wurster, M., Kreisel, H. and Lindequist, U. 2005. Antimicrobial, cytotoxic and antioxidant activity of selected *Basidiomycetes* from Yemen. *Pharm.* **60**: 776-780.
- Al-Fatimi, M., Jülich, W.D., Jansen, R. and Lindequist, U. 2006. Bioactive components of the traditionally used mushroom *Podaxis pistillaris*. *Evid. Based Complement. Alternat. Med.* **3**: 87-92.
- Allbutt, A.D., Ayer, W.A., Brodie, H.J., Johri, B.N. and Taube, H. 1971. Cyathin, a new antibiotic complex produced by *Cyathus helenae*. *Can. J. Microbiol.* **17**: 1401-1407.
- Almendros, G., Martín, F., González-Vila, F.J. And Martínez, A.T. 1987. Melanins and lipids in *Lycoperdon perlatum* fruit bodies. *Trans. Br. Mycol. Soc.* **89**: 533-537.
- Amdekar, S., Roy, P., Singh, V., Kumar, A., Singh, R. and Sharma, P. 2012. Anti-Inflammatory Activity of *Lactobacillus* on carrageenan-induced paw edema in male wistar rats. *Int. J. Inflamm.* **2012**: 752015.
- Aniekemabasi, U.I., Basil, N.I. and Uwemobong, B.A. 2018. Physicochemical properties of starches from *Scleroderma citrinum* and *Pleurotus ostreatus*. *Pharm. Chem. J.* **5**: 191-195.
- Anilkumar, M. 2010. Ethnomedicinal Plants as Anti-inflammatory and Analgesic Agents. In: *Ethnomedicine: A Source of Complementary Therapeutics*. (Ed.: Chattopadhyay, D.), Research Signpost, pp. 267-294.
- Anke, T., Oberwinkler, F., Steglich, W. and Hofle, G. 1977. The striatins-new antibiotics from the basidiomycete *Cyathus striatus* (Huds. ex Pers.) Willd. *J. Antibiot.* **30**: 221-225.
- Aulinger, K., Besl, H., Spiteller, P., Spiteller, M. and Steglich, W. 2001. Melanocrocins, a polyene pigment from *Melanogaster broomeianus* (Basidiomycetes). *Z. Für Naturforschung C.* **56**: 495-498.
- Ayer, W.A. and Taube, H. 1972. Metabolites of *Cyathus helenae*. cyathin A3 and allocyathin B3, members of a new group of diterpenoids. *Tetrahedron Lett.* **13**: 1917-1920.
- Ayer, W.A. and Singer, P.P. 1980. Phenolic metabolites of the bird's nest fungus *Nidula niveo-tomentosa*. *Phytochemistry* **19**: 2717-2721.
- Barros, L., Baptista, P., Correia, D.M., Casal, S., Oliveira, B. and Ferreira, I.C.F.R. 2007. Fatty acid and sugar compositions, and nutritional value of five wild edible mushrooms from Northeast Portugal. *Food Chem.* **105**: 140-145.
- Barros, L., Cruz, T., Baptista, P., Estevinho, L.M. and Ferreira, I.C.F.R. 2008. Wild and commercial mushrooms as source of nutrients and nutraceuticals. *Food Chem. Toxicol.* **46**: 2742-2747.
- Barros, L., Dueñas, M., Ferreira, I.C.F.R., Baptista, P. and Santos-Buelga, C. 2009. Phenolic acids determination by HPLC/DAESI/MS in sixteen different Portuguese wild mushrooms species. *Food Chem. Toxicol.* **47**: 1076-1079.
- Bayman, P., Ritchey, S.D. and Bennett, J.W. 1995. Fungal

- interactions with the explosive RDX (hexahydro-1,3,5-trinitro-1,3,5-triazine). *J. Ind. Microbiol.* **15**: 418-423.
- Bedford, M.R. 2000. Exogenous enzymes in monogastric nutrition their current value and future benefits. *Anim. Feed Sci. Technol.* **86**: 1-13.
- Berkeley, M.J. 1867. On some new fungi from Mexico. *Bot. J. Linn. Soc.* **9**: 423-425.
- Bisht, K., Tampe, J., Shing, C., Bakrania, B., Winearls, J., Fraser, J., Wagner, K.H. and Bulmer, A.C. 2014. Endogenous tetrapyrroles influence leukocyte responses to lipopolysaccharide in human blood: pre-clinical evidence demonstrating the anti-inflammatory potential of biliverdin. *J. Clin. Cell. Immunol.* **5**: 1000218.
- Biswas, G., Sarkar, S. and Acharya, K. 2010. Free radical scavenging and anti-inflammatory activities of the extracts of *Astraeus hygrometricus* (Pers.) Morg. *Lat. Am. J. Pharm.* **29**: 549-553.
- Biswas, G., Sarkar, S. and Acharya, K. 2011. Hepatoprotective activity of the ethanolic extract of *Astraeus hygrometricus* (Pers.) Morg. *Dig. J. Nanomater. Biostructures* **6**: 637-641.
- Blackwell, M. and Spatafora, J.W. 2004. Fungi and their Allies. In: *Biodiversity of Fungi: Inventory and Monitoring Methods*. (Eds.: Mueller, G.M., Bills, G.F., Foster, M.S.). Academic Press, pp. 721.
- Bon, M., Wilkinson, J. and Ovenden, D. 1987. *The Mushrooms and Toadstools of Britain and North-Eastern Europe*. Hodder & Stoughton, London. pp. 352.
- Brodie, H. 1975. *The Bird's Nest Fungi*. University of Toronto Press, Canada. pp. 216.
- Caleja, C., Barros, L., Antonio, A.L., Oliveira, M.B.P.P. and Ferreira, I.C.F.R. 2017. A comparative study between natural and synthetic antioxidants: Evaluation of their performance after incorporation into biscuits. *Food Chem.* **216**: 342-346.
- Chakraborty, I., Mondal, S., Pramanik, M., Rout, D. and Islam, S.S. 2004. Structural investigation of a water-soluble glucan from an edible mushroom, *Astraeus hygrometricus*. *Carbohydr. Res.* **339**: 2249-2254.
- Chang, T.S. 2009. An updated review of tyrosinase inhibitors. *Int. J. Mol. Sci.* **10**: 2440-2475.
- Chang, S.T. and Miles, P.G. 2004. *Mushrooms: Cultivation, Nutritional Value, Medicinal Effect, and Environmental Impact*. CRC Press. pp. 480.
- Chhabra, M., Mishra, S. and Sreekrishnan, T.R. 2008. Mediator-assisted decolorization and detoxification of textile dyes/dye mixture by *Cyathus bulleri* laccase. *Appl. Biochem. Biotechnol.* **151**: 587-598.
- Chittaragi, A., Naika, R., Ashwini, H. and Nagaraj, K. 2013. Antibacterial potential of *Geastrum triplex* Jungh. against plant and human pathogens. *Int. J. Pharmtech. Res.* **5**: 1456-1464.
- Chittaragi, A., Naika, R. and Vinayaka, K.S. 2014. Nutritive value of few wild mushrooms from the Western Ghats of Shivamogga district, Karnataka, India. *Asian J. Pharm. Clin. Res.* **7**: 50-53.
- Coetze, J.C. and Wyk, A. van. 2009. The genus *Calvatia* (*Gasteromycetes*, *Lycoperdaceae*): A review of its ethnomycology and biotechnological potential. *Afr. J. Biotechnol.* **8**: 6007-6015.
- Cohen, R., Persky, L. and Hadar, Y. 2002. Biotechnological applications and potential of wood-degrading mushrooms of the genus *Pleurotus*. *Appl. Microbiol. Biotechnol.* **58**: 582-594.
- Colak, A., Faiz, Ö. and Sesli, E. 2009. Nutritional composition of some wild edible mushrooms. *Turk. J. Biochem.* **34**: 25-31.
- Coletto, M.B. 2006. Antibiotic activity in *Basidiomycetes*. XIV. Antibacterial and antifungal activity of some new recently isolated strains. *Allionia* **40**: 33-37.
- Coletto, M. and Lelli, P. 1998. Antibiotic activity in *Basidiomycetes*. XII. Antibacterial and antifungal activity of 32 new strains. *Allionia* **36**: 89-92.
- Crisan, E.V. and Sands, A. 1978. Nutritional Value. In: *The Biology and Cultivation of Edible Mushrooms*. (Eds.: Chang, S.T. and Hayes, W.A.). Academic Press, pp. 137-168.
- Davis, R.M., Sommer, R. and Menge, J.A. 2012. Field guide to mushrooms of Western North America. University of California Press, California. pp. 459.
- Deng, C., Shang, J., Fu, H., Chen, J., Liu, H. and Chen, J. 2016. Mechanism of the immunostimulatory activity by a polysaccharide from *Dictyophora indusiata*. *Int. J. Biol. Macromol.* **91**: 752-759.
- Devi, K., Brahma, J. and Shrivastava, K. 2016. Documentation of four hitherto unreported wild edible macro fungi from Chirang district of Assam, north-east India. *Int. J. Conserv. Sci.* **7**: 709-718.
- Diallo, D., Sogn, C., Samaké, F.B., Paulsen, B.S., Michaelsen, T.E. and Keita, A. 2002. Wound healing plants in Mali, the Bamako region. An ethnobotanical survey and complement fixation of water extracts from selected plants. *Pharm. Biol.* **40**: 117-128.
- Dickinson, C. and Lucas, J. 1979. *The Encyclopedia of Mushrooms*. GP Putnam's Sons, New York. pp. 280.
- Dickinson, C.H. and Lucas, J.A. 1982. *VNR Color Dictionary of Mushrooms*. Van Nostrand Reinhold.

- Dore, C.M.P.G., Azevedo, T.C.G., de Souza, M.C.R., Rego, L.A., de Dantas, J.C.M., Silva, F.R.F., Rocha, H.A.O., Baseia, I.G. and Leite, E.L. 2007. Anti-inflammatory, antioxidant and cytotoxic actions of β -glucan-rich extract from *Geastrum saccatum* mushroom. *Int. Immunopharmacol.* **7**: 1160-1169.
- Doshi, A. and Bohra, B. 2000. Studies on the gastromycetous fungus *Phellorinia inquinans* Berk. *Final Project Report*, ICAR project PG/Gr.II/ICAR/14/95-96.
- Doshi, A., Bohra, B. and Sharma, S.S. 1999. Studies on basidiospore germination of *Phellorinia inquinans* Berk.. *Micol. Ital.* **28**: 33-37.
- Dulger, B. 2005. Antimicrobial activity of ten *Lycoperdaceae*. *Fitoterapia* **76**: 352-354.
- Ek, M., Ljungquist, P.O. and Stenström, E. 1983. Indole-3-acetic acid production by mycorrhizal fungi determined by gas chromatography-mass spectrometry. *New Phytol.* **94**: 401-407.
- Erkel, G., Becker, U., Anke, T. and Sterner, O. 1996. Nidulal, a novel inducer of differentiation of human promyelocytic leukemia cells from *Nidula candida*. *J. Antibiot.* **49**: 1189-1195.
- Escobar, G.A., McCabe, D.E. and Harpel, C.W. 1976. *Limnoperdon*, a floating gasteromycete isolated from marshes. *Mycologia* **68**: 874-880.
- Farrington, E.A. 1999. *Lectures on Clinical Materia Medica*. B. Jain Publishers (P) Limited, New Delhi, India. pp. 1037.
- Fiasson, J.L. and Petersen, R.H. 1973. Carotenes in the fungus *Clathrus ruber* (*Gasteromycetes*). *Mycologia* **65**: 201-203.
- Garbe, D. 2000. Cinnamic Acid. In: *Ullmann's Encyclopedia of Industrial Chemistry*. Wiley-VCH Verlag, 193-195.
- García-Lafuente, A., Guillamón, E., Villares, A., Rostagno, M.A. and Martínez, J.A. 2009. Flavonoids as anti-inflammatory agents: implications in cancer and cardiovascular disease. *Inflamm. Res.* **58**: 537-552.
- Gehlot, P. 2016. Edible *Gastromycetes* mushrooms of Thar desert: new economic source for rural upliftment. *Mushroom Res.* **25**: 55-56.
- Gehlot, P. and Singh, S. 2015. Diversity of gastromycetous flora in Indian Thar Desert. *Indian J. Trop. Biodivers.* **23**: 74-77.
- Gehlot, P. and Singh, S. 2016. Ethno-mycological studies of wild edible mushroom *Podaxis pistilaris* (L. ex Pers.) Fr.: A mini review. *J. Trop. For.* **32**: 58-68.
- Gehlot, S. and Gehlot, P. 2020. Socio-ethnomycological validation of Maru khumbhi (*Phellorinia herculeana*) occurring in Indian Thar Desert. *NEHU J.* **18**: 22-27.
- Gehlot, P., Pathak, R. and Singh, S.K. 2016. *Phellorinia*: a under exploited mushroom. *Indian J. Trop. Biodivers.* **24**: 17.
- Gehlot, P., Sharma, K., Solanki, D.S., Kumar, S., Parihar, K., Pathak, R. and Singh, S.K. 2018. Identification and characterization of antibacterial bioactive compounds from an edible gasteroid mushroom *Phellorinia herculeana*. *J. Mycol. Plant Pathol.* **48**: 178-182.
- Gehlot, P., Solanki, D.S., Kumar, S., Parihar, K., Tak, A., Pathak, R. and Singh, S.K. 2020. A new record of gasteroid fungus *Broomeia congregata* Berk. from Great Indian Thar Desert, India. *Indian Phytopathol.* **73**: 111-115.
- Gill, M. and Lally, D.A. 1985. A naphthalenoid pulvinic acid derivative from the fungus *Pisolithus tinctorius*. *Phytochemistry* **24**: 1351-1354.
- Gogoi, G. and Vipin, P. 2015. Diversity of gasteroid fungi (Basidiomycota) in Hollongapar Gibbon Wildlife Sanctuary, Jorhat, Assam, India. *Curr. Res. Environ. Appl. Mycol.* **5**: 202-212.
- Gruber, G. and Steglich, W. 2007. Calostomal, a polyene pigment from the gasteromycete *Calostoma cinnabarinum* (*Boletales*). *Z. Für Naturforschung B* **62**: 129-131.
- Gülçin, İ., Oktay, M., Küfrevioğlu, Ö.İ. and Aslan, A. 2002. Determination of antioxidant activity of lichen *Cetraria islandica* (L) Ach. *J. Ethnopharmacol.* **79**: 325-329.
- Habtemariam, S. 2019. The chemistry, pharmacology and therapeutic potential of the edible mushroom *Dictyophora indusiata* (Vent ex. Pers.) Fischer (syn. *Phallus indusiatus*). *Biomedicines* **7**: 98.
- Halliwell, B. and Gutteridge, J.M.C. 2015. *Free Radicals in Biology and Medicine*. Oxford University Press, Oxford. pp. 944.
- Halpern, G.M. 2007. *Healing Mushrooms*. Square One Publishers, New York. pp. 184.
- Han, S., Ma, C., Hu, M., Wang, Y., Ma, F., Tao, N. and Qin, Z. 2017. A polysaccharide from *Dictyophora indusiata* inhibits the immunosuppressive function of cancer-associated fibroblasts. *Cell Biochem. Funct.* **35**: 414-419.
- Hao, J., Zhang, G., Han, H., Wang, D. and He, Y. 2008. Study on the extracted method of *Dictyophora indusiata* Fischer and its antimicrobial action. *Sci. Technol. Food Ind.* **29**: 123-127.
- Hara, C., Kiho, T., Tanaka, Y. and Ukai, S. 1982. Anti-inflammatory activity and conformational behavior of a branched (1 \rightarrow 3)- β -d-glucan from an alkaline extract of *Dictyophora indusiata* Fisch. *Carbohydr. Res.* **110**: 77-87.

- Hasan, M., Khatun, M., Sajib, M., Rahman, M., Rahman, M., Roy, M., Miah, M. and Ahmed, K. 2015. Effect of wheat bran supplement with sugarcane bagasse on growth, yield and proximate composition of pink oyster mushroom (*Pleurotus djamor*). *Am. J. Food Sci. Technol.* **3**: 150-157.
- Hettiarachchy, N.S., Glenn, K.C., Gnanasambandam, R. and Johnson, M.G. 1996. Natural antioxidant extract from fenugreek (*Trigonella foenum-graceum*) for ground beef patties. *J. Food Sci.* **61**: 516-519.
- Hibbett, D.S. and Binder, M. 2001. Evolution of marine mushrooms. *Biol. Bull.* **201**: 319-322.
- Hobbs, C. 1995. *Medicinal Mushrooms: An Exploration of Tradition, Healing, and Culture*. Botanica Press, USA. pp. 251.
- Holliday, J. and Soule, N. 2001. Spontaneous female orgasms triggered by the smell of a newly found tropical *Dictyophora* Desv. species. *Int. J. Med. Mushrooms* **3**.
- Hong-Jun, S., Li-Zhen, F., Wan-Qiu, Y., Fei, W. and Ji-Kai, L. 2007. A new Steryl Ester from the culture mycelia of the basidiomycete *Astraeus hygrometricus* (*Astraceae*). *Plant Divers.* **29**: 371-374.
- Huang, M., Chen, X., Tian, H., Sun, B. and Chen, H. 2011. Isolation and identification of antibiotic albaflavenone from *Dictyophora indusiata* (Vent. Pers.) Fischer. *J. Chem. Res.* **35**: 659-660.
- Iftekhhar, A.F.M.H., Choudhry, Z.K., Khan, M.I. and Saleh, A.A. 2011. Comparative study of antibacterial activity of wood-decay fungi and antibiotics. *Bangladesh J. Pharmacol.* **6**: 14-17.
- Iwalokun, A.B., Usen, A.U., Otunba, A.A. and Olukoya, K.D. 2007. Comparative phytochemical evaluation, antimicrobial and antioxidant properties of *Pleurotus ostreatus*. *Afr. J. Biotechnol.* **6**: 1732-1739.
- Jain, J.P. 1997. *Highlights of Homoeopathic Materia Medica*. B. Jain Publishers Pvt. Limited, New Delhi, India. pp. 516.
- Jandaik, C.L. 1977. Nutritive value of *Phellorina inquinans* Berk. - an edible gasteromycete (India). *Indian J. Mushrooms* **3**: 58.
- Jasalavich, C.A., Ostrofsky, A. and Jellison, J. 2000. Detection and identification of decay fungi in spruce wood by restriction fragment length polymorphism analysis of amplified genes encoding rRNA. *Appl. Environ. Microbiol.* **66**: 4725-4734.
- Jonathan, G., Odebode, C. and Bawo, D. 2008. Studies on collection and proximate composition of *Phallus indusiatus* (Vent. ex. Pers), a Nigerian higher fungus. *World J. Agric. Sci.* **4**: 18-22.
- Julich, W. 1981. *Higher Taxa of Basidiomycetes*. J. Cramer, Vaduz, Liechtenstein. pp. 485.
- Kakumyan, P. and Matsui, K. 2009. Characterization of volatile compounds in *Astraeus* spp. *Biosci. Biotechnol. Biochem.* **73**: 2742-2745.
- Kakumyan, P., Suwannarach, N., Kumla, J., Saichana, N., Lumyong, S. and Matsui, K. 2020. Determination of volatile organic compounds in the stinkhorn fungus *Pseudocolus fusiformis* in different stages of fruiting body formation. *Mycoscience* **61**: 65-70.
- Kalač, P. 2016. *Edible Mushrooms*. Academic Press, 236.
- Kang, H.S., Jun, E.M., Park, S.H., Heo, S.J., Lee, T.S., Yoo, I.D. and Kim, J.P. 2007. Cyathusals A, B, and C, antioxidants from the fermented mushroom *Cyathus stercoreus*. *J. Nat. Prod.* **70**: 1043-1045.
- Kawagishi, H., Ishiyama, D., Mori, H., Sakamoto, H., Ishiguro, Y., Furukawa, S. and Li, J. 1997. Dictyophorines A and B, two stimulators of NGF-synthesis from the mushroom *Dictyophora indusiata*. *Int. J. Plant Biochem. Mol. Biol.* **45**: 1203-1205.
- Ker, Y.B., Chen, K.C., Peng, C.C., Hsieh, C.L. and Peng, R.Y. 2011. Structural characteristics and antioxidative capability of the soluble polysaccharides present in *Dictyophora indusiata* (Vent. ex. Pers.) Fish. *Phallaceae. Evid. Based Complement. Alternat. Med.* **2011**: 396013.
- Kıvrak, İ., Kıvrak, Ş. and Harmandar, M. 2014. Free amino acid profiling in the giant puffball mushroom (*Calvatia gigantea*) using UPLCMS/MS. *Food Chem.* **158**: 88-92.
- Kong, X., Kasapis, S., Bao, J. and Corke, H. 2009. Effect of gamma irradiation on the thermal and rheological properties of grain amaranth starch. *Radiat. Phys. Chem.* **78**: 954-960.
- Kovacs, B., Béni, Z., Dékány, M., Bózsity, N., Zupko, I., Hohmann, J. and Vanyolos, A. 2018. Isolation and structure determination of antiproliferative secondary metabolites from the potato earthball mushroom, *Scleroderma bovista* (*Agaricomycetes*). *Int. J. Med. Mushrooms* **20**: 411-418.
- Krzyczkowski, W., Malinowska, E. and Herold, F. 2008. Budowa, właściwości lecznicze i biosynteza diterpenoidów kjetanowych. *Biotechnologia* **1**: 146-167.
- Kumar, M., Harsh, N.S.K., Prasad, R. and Pandey, V.V. 2017. An ethnomycological survey of Jaunsar, Chakrata, Dehradun, India. *J. Threat. Taxa* **9**: 10717-10725.
- Kuznecov, G., Jegina, K., Kuznecovs, S. and Kuznecovs, I. 2007. P151 *Phallus impudicus* in thromboprophylaxis in breast cancer patients undergoing chemotherapy and hormonal treatment. *The Breast* **16**: S56.

- Kyselova, Z., Stefek, M. and Bauer, V. 2004. Pharmacological prevention of diabetic cataract. *J. Diabetes Complications* **18**: 129-140.
- Læssøe, T. and Spooner, B. 1994. The uses of 'Gasteromycetes'. *Mycologist* **8**: 154-159.
- Lam, Y.W., Ng, T.B. and Wang, H.X. 2001. Antiproliferative and antimutagenic activities in a peptide from puffball mushroom *Calvatia caelata*. *Biochem. Biophys. Res. Commun.* **289**: 744-749.
- Lightfoot, J. 1789. *Flora Scotica: or, a Systematic Arrangement, in the Linnaean Method, of the Native Plants of Scotland and the Hebrides*. R. Faulder, London. pp. 644.
- Liu, J.K. 2005. N-containing compounds of macromycetes. *Chem. Rev.* **105**: 2723-2744.
- Liu, B. and Bau, Y. 1980. *Fungi Pharmacopoeia (Sinica)*. Kinoko Company, Oakland. pp. 297.
- Liu, Y.J. and Zhang, K.Q. 2004. Antimicrobial activities of selected *Cyathus* species. *Mycopathologia* **157**: 185-189.
- Liu, F., Ooi, V.E.C. and Chang, S.T. 1997. Free radical scavenging activities of mushroom polysaccharide extracts. *Life Sci.* **60**: 763-771.
- Liu, Y.T., Sun, J., Luo, Z.Y., Rao, S.Q., Su, Y.J., Xu, R.R. and Yang, Y.J. 2012. Chemical composition of five wild edible mushrooms collected from Southwest China and their antihyperglycemic and antioxidant activity. *Food Chem. Toxicol.* **50**: 1238-1244.
- Łopusiewicz, Ł., Jędra, F. and Mizielińska, M. 2018. New Poly (lactic acid) active packaging composite films incorporated with fungal melanin. *Polymers* **10**: 386.
- Maiti, S., Bhutia, S.K., Mallick, S.K., Kumar, A., Khadgi, N. and Maiti, T.K. 2008. Antiproliferative and immunostimulatory protein fraction from edible mushrooms. *Environ. Toxicol. Pharmacol.* **26**: 187-191.
- Malik, A.R., Wani, A.H., Bhat, M.Y. and Parveen, S. 2017. Ethnomycological knowledge of some wild mushrooms of northern districts of Jammu and Kashmir, India. *Asian J. Pharm. Clin. Res.* **10**: 399-405.
- Mallick, S.K., Maiti, S., Bhutia, S.K. and Maiti, T.K. 2010. Antitumor properties of a heteroglucan isolated from *Astraeus hygrometricus* on Dalton's lymphoma bearing mouse. *Food Chem. Toxicol.* **48**: 2115-2121.
- Mallick, S.K., Maiti, S., Bhutia, S.K. and Maiti, T.K. 2011. Activation of RAW 264.7 cells by *Astraeus hygrometricus*-derived heteroglucan through MAP kinase pathway. *Cell Biol. Int.* **35**: 617-621.
- Manna, S., Ray, D. and Roy, A. 2014. Tribal relation to spatio-temporal variation of wild mushrooms in eastern lateritic part of India. *Ethnobot. Res. Appl.* **12**: 015-024.
- Mao, X. 2000. *The Macrofungi in China*. Henan Science and Technology Press, Zhengzhou, China. pp. 719.
- Matsumoto, K., Nagashima, K., Kamigauchi, T., Kawamura, Y., Yasuda, Y., Ishii, K., Uotani, N., Sato, T., Nakai, H. and Terui, Y. 1995. Salfredins, new aldose reductase inhibitors produced by *Crucibulum* sp. RF-3817. I. Fermentation, isolation and structures of salfredins. *J. Antibiot.* **48**: 439-446.
- Mau, J.L., Lin, H.C. and Song, S.F. 2002. Antioxidant properties of several specialty mushrooms. *Food Res. Int.* **35**: 519-526.
- McIlvaine, C. and Macadam, R.K. 1900. *One Thousand American Fungi*. Bowen-Merrill Company, Indianapolis. pp. 704.
- McKnight, K.H. and McKnight, V.B. 1998. *A Field Guide to Mushrooms: North America*. Houghton Mifflin Harcourt, Boston, USA. pp. 429.
- Meuninck, J. 2017. *Foraging Mushrooms Oregon: Finding, Identifying, and Preparing Edible Wild Mushrooms*. Falcon Guides, Guilford, Connecticut. pp. 168.
- Miller, O.K. and Miller, H.H. 2006. *North American Mushrooms a Field Guide to Edible and Inedible Fungi*. Falcon Guides, Guilford, Connecticut. pp. 592.
- Mohan, V., Nivea, R. and Menon, S. 2015. Evaluation of ectomycorrhizal fungi as potential bio-control agents against selected plant pathogenic fungi. *J. Acad. Ind. Res.* **3**: 408-412.
- Mridu and Atri, N.S. 2017. Nutritional and nutraceutical characterization of three wild edible mushrooms from Haryana, India. *Mycosphere* **8**: 1035-1043.
- Mueller, G.M. and Schmit, J.P. 2007. Fungal biodiversity: what do we know? What can we predict? *Biodivers. Conserv.* **16**: 15.
- Muhsin, T.M., Abass, A.F. and Al-Habeeb, E.K. 2012. *Podaxis pistillaris* (Gasteromycetes) from the desert of southern Iraq, an addition to the known mycota of Iraq. *J. Basrah Res. Sci.* **38**: 29-35.
- Nedelcheva, D., Antonova, D., Tsvetkova, S., Marekov, I., Momchilova, S., Nikolova-Damyanova, B. and Gyosheva, M. 2007. TLC and GC-MS probes into the fatty acid composition of some *Lycoperdaceae* mushrooms. *J. Liq. Chromatogr. Relat. Technol.* **30**: 2717-2727.
- Newman, J.M. 1999. Bamboo mushrooms. *Flavor Fortune* **6**: 25-30.
- Ogbole, O.O., Nkumah, A.O., Linus, A.U. and Falade, M.O.

2019. Molecular identification, *in vivo* and *in vitro* activities of *Calvatia gigantea* (macro-fungus) as an antidiabetic agent. *Mycology* **10**: 166-173.
- Olennikov, D.N., Tankhaeva, L.M. and Agafonova, S.V. 2011. Antioxidant components of *Laetiporus sulphureus* (Bull.: Fr.) Murr. fruit bodies. *Appl. Biochem. Microbiol.* **47**: 419-425.
- Oso, B.A. 1975. Mushrooms and the Yoruba People of Nigeria. *Mycologia* **67**: 311-319.
- Pala, S.A., Wani, A.H. and Bhat, M.Y. 2013. Ethnomycological studies of some wild medicinal and edible mushrooms in the Kashmir Himalayas (India). *Int. J. Med. Mushrooms* **15**: 211-220.
- Panda, M., Thatoi, H. and Tayung, K. 2017. Antimicrobial potentials of spore culture of *Geastrum* sp., a rare wild edible mushroom of Similipal Biosphere Reserve, Odisha, India against some significant human pathogens. *Int. J. Adv. Res.* **5**: 1508-1516.
- Panwar, C.H. and Purohit, D.K. 2002. Antimicrobial activities of *Podaxis pistillaris* and *Phellorinia inquinans* against *Pseudomonas aeruginosa* and *Proteus mirabilis*. *Mushroom Res.* **11**: 43-44.
- Pavithra, M., Sridhar, K.R. and Greeshma, A.A. 2018. Nutritional Quality Attributes of Edible Gasteroid Wild Mushroom *Astraeus hygrometricus*. In: *Fungi and their Role in Sustainable Development: Current Perspectives*. (Ed.: Gehlot, P. and Singh, J.). Springer Nature, 367-382.
- Pérez-Silva, E., Herrera, T. and Medina-Ortiz, A.J. 2015. *Mycenastrum corium* and gastrointestinal mycetism in México. *Mycotaxon* **130**: 641-645.
- Phutela, R., Kaur, H. and Sodhi, H. 1998. Physiology of an edible gasteromycete, *Podaxis pistillaris* (Lin. ex. Pers) Fr. *J. Mycol. Plant Pathol.* **28**: 31-37.
- Pujol, V., Seux, V. and Villard, J. 1990. Research of antifungal substances secreted by higher fungi in culture. *Ann. Pharm. Fr.* **48**: 17-22.
- Rai, B.K., Ayachi, S.S. and Rai, A. 1993. A note on ethno-myco-medicines from central India. *Mycologist* **7**: 192-193.
- Ramesh, C.H. and Pattar, M.G. 2010. Antimicrobial properties, antioxidant activity and bioactive compounds from six wild edible mushrooms of Western Ghats of Karnataka, India. *Pharmacogn. Res.* **2**: 107-112.
- Ramsbottom, J. 1953. Mushrooms and toadstools. *Proc. Nutr. Soc.* **12**: 39-44.
- Rinaldi, A. and Tyndalo, V. 1985. *Complete Book of Mushrooms*. Crescent, New York. pp. 310.
- Roberts, P. and Evans, S. 2011. *The Book of Fungi*. The University of Chicago Press, Chicago, US. pp. 656.
- Rolfe, R.T. and Rolfe, F.W. 1925. The romance of the fungus world. Chapman & Hall. Limited, London. pp. 309.
- Sano, M., Yoshino, K., Matsuzawa, T. and Ikekawa, T. 2002. Inhibitory effects of edible higher Basidiomycetes mushroom extracts on mouse type IV allergy. *Int. J. Med. Mushrooms* **4**: 37-41.
- Sarac, N., Alli, H., Baygar, T. and Ugur, A. 2019. *In vitro* anticoagulant and antiinflammatory activities of *Geastrum fimbriatum* Fr., namely as earthstar fungus. *Int. J. Second. Metab.* **6**: 19.
- Scarpa, G.F. 2004. Medicinal plants used by the Criollos of Northwestern Argentine Chaco. *J. Ethnopharmacol.* **91**: 115-135.
- Sethuraman, A., Akin, D.E. and Eriksson, K.E.L. 1999. Production of ligninolytic enzymes and synthetic lignin mineralization by the bird's nest fungus *Cyathus stercoreus*. *Appl. Microbiol. Biotechnol.* **52**: 689-697.
- Sevindik, M., Akgul, H., Akata, I. and Selamoglu, Z. 2017. *Geastrum pectinatum* as an alternative antioxidant source with some biochemical analysis. *Med. Mycol.* **3**: 25.
- Sharma, Y. and Doshi, A. 1996. Some studies on an edible wild fungus *Phellorinia inquinans* Berk. In Rajasthan, India. *Mushroom Res.* **5**: 51-53.
- Sharma, V.K., Choi, J., Sharma, N., Choi, M. and Seo, S.Y. 2004. In vitro anti-tyrosinase activity of 5-(hydroxymethyl)-2-furfural isolated from *Dictyophora indusiata*. *Phytother. Res.* **18**: 841-844.
- Sharma, V., Singh, M., Kumar, S., Kamal, S. and Singh, R. 2015. Phylogene and physiology of *Phellorinia* spp.: a delicacy of Indian desert. *Int. Res. J. Nat. Appl. Sci.* **2**: 1-17.
- Sims, K.P., Watling, R. and Jeffries, P. 1995. A revised key to the genus *Scleroderma*. *Mycotaxon* **56**: 403-420.
- Singh, S., Doshi, A., Yadav, M. and Kamal, S. 2006. Molecular characterization of specialty mushrooms of western Rajasthan, India. *Curr. Sci.* **91**: 1225-1230.
- Singh, P., Singh, A., D'Souza, L.M., Roy, U. and Singh, S.M. 2012. Chemical constituents and antioxidant activity of the Arctic mushroom *Lycoperdon molle* Pers. *Polar Res.* **31**: 17329.
- Smith, A.H. and Weber, N.S. 1980. *The Mushroom Hunter's Field Guide*. University of Michigan Press, Michigan, US. pp. 324.
- Solano, F. 2014. Melanins: Skin pigments and much moretypes, structural models, biological functions, and formation routes. *New J. Sci.* **2014**: 498-276.
- Soytong, K., Sibounnavong, P., Kanokmedhakul, K. and Kanokmedhakul, S. 2014. Biological active compounds of *Scleroderma citrinum* that inhibit

- plant pathogenic fungi. *J. Agric. Technol.* **10**: 79-86.
- Srivastava, S.K., Ramana, K.V. and Bhatnagar, A. 2005. Role of aldose reductase and oxidative damage in diabetes and the consequent potential for therapeutic options. *Endocr. Rev.* **26**: 380-392.
- Stanikunaite, R., Radwan, M.M., Trappe, J.M., Fronczek, F. and Ross, S.A. 2008. Lanostane-type triterpenes from the mushroom *Astraeus pteridis* with antituberculosis activity. *J. Nat. Prod.* **71**: 2077-2079.
- Suay, I., Arenal, F., Asensio, F.J., Basilio, A., Angeles Cabello, M., Teresa Díez, M., García, J.B., González del Val, A., Gorrochategui, J., Hernández, P., Peláez, F. and Francisca Vicente, M. 2000. Screening of Basidiomycetes for antimicrobial activities. *Antonie Van Leeuwenhoek* **78**: 129-140.
- Tan, K.H., Sihanonth, P. and Todd, R.L. 1978. Formation of humic acid like compounds by the ectomycorrhizal fungus, *Pisolithus tinctorius*. *Soil Sci. Soc. Am. J.* **42**: 906-908.
- Thangaraj, R., Raj, S. and Renganathan, K. 2017. Wound healing effect of King Alferd's mushroom (*Daldinia concentrica*) used by tribes of Sirumalai hills, Tamilnadu, India. *Int. J. Pharm. Pharm. Sci.* **9**: 161-164.
- Thatoi, H. and Singdevsachan, S.K. 2014. Diversity, nutritional composition and medicinal potential of Indian mushrooms: A review. *Afr. J. Biotechnol.* **13**: 523-545.
- Tibuhwa, D.D. 2012. Folk taxonomy and use of mushrooms in communities around Ngorongoro and Serengeti National Park, Tanzania. *J. Ethnobiol. Ethnomed.* **8**: 36.
- Trierveiler-Pereira, L. and Baseia, I.G. 2009. A checklist of the Brazilian gasteroid fungi (*Basidiomycota*). *Mycotaxon* **108**: 441-444.
- Tsantrizos, Y.S., Kope, H.H., Fortin, J.A. and Ogilvie, K.K. 1991. Antifungal antibiotics from *Pisolithus tinctorius*. *Phytochemistry* **30**: 1113-1118.
- Ukai, S., Hara, C., Kiho, T. and Hirose, K. 1980. Polysaccharides in Fungi. V. Isolation and characterization of a mannan from aqueous ethanol extract of *Dictyophora indusiata* Fisch. *Chem. Pharm. Bull. (Tokyo)* **28**: 2647-2652.
- Ukai, S., Kiho, T., Hara, C., Morita, M., Goto, A., Imaizumi, N. and Hasegawa, Y. 1983. Polysaccharides in fungi. XIII. antitumor activity of various polysaccharides isolated from *Dictyophora indusiata*, *Ganoderma japonicum*, *Cordyceps cicadae*, *Auricularia auricula-judae*, and *Auricularia* species. *Chem. Pharm. Bull. (Tokyo)* **31**: 741-744.
- Vasdev, K., Kuhad, R.C. and Saxena, R.K. 1995. Decolorization of triphenylmethane dyes by the bird's nest fungus *Cyathus bulleri*. *Curr. Microbiol.* **30**: 269-272.
- Vasquez-Davila, M.A. 2017. Current and potential use of the desert fungus *Podaxis pistillaris* (L.) Fr. (*Agaricaceae*). *J. Bacteriol. Mycol.* **5**: 307-309.
- Vishwakarma, P., Tripathi, N. and Singh, P. 2017. A checklist of macrofungi of Gorakhpur District, UP India. *Curr. Res. Environ. Appl. Mycol.* **7**: 109-120.
- Wang, W., Liu, H., Zhang, Y., Feng, Y., Yuan, F., Song, X., Gao, Z., Zhang, J., Song, Z. and Jia, L. 2019. Antihyperlipidemic and hepatoprotective properties of alkali- and enzyme-extractable polysaccharides by *Dictyophora indusiata*. *Sci. Rep.* **9**: 142-66.
- Wei, S. 2005. *Dictyophora indusiata* and its nutritive values. *Chin. Food Nutr.* **4**: 15-16.
- Wicklow, D.T., Detroy, R.W. and Jessee, B.A. 1980. Decomposition of lignocellulose by *Cyathus stercoreus* (Schw.) de Toni NRRL 6473, a "white rot" fungus from cattle dung. *Appl. Environ. Microbiol.* **40**: 169-170.
- Winner, M., Giménez, A., Schmidt, H., Sontag, B., Steffan, B. and Steglich, W. 2004. Unusual pulvinic acid dimers from the common fungi *Scleroderma citrinum* (common earthball) and *Chalciporus piperatus* (peppery bolete). *Angew. Chem. Int. Ed.* **43**: 1883-1886.
- Xianling, G., Rensen, Z., Shiming, L., Chongren, Y. and Qingan, Z. 2005. Chemical constituents from the fruit bodies of *Scleroderma polyrhizum*. *Nat. Prod. Res. Dev.* **17**: 431-433.
- Yahaya, Y.A. and Don, M.M. 2012. Evaluation of *Trametes lactinea* extracts on the inhibition of hyaluronidase, lipoxigenase and xanthine oxidase activities in vitro. *J. Phys. Sci.* **23**: 1-15.
- Yin, J. and Zhou, L. 2008. Analysis of nutritional components of 4 kinds of wild edible fungi in Yunnan. *Food Res. Dev.* **29**: 133-136.
- Ying, C. 1987. *Icons of Medicinal Fungi from China*. Science Press, Beijing, China. pp. 575.
- Yun, W. and Hall, I.R. 2004. Edible ectomycorrhizal mushrooms: challenges and achievements. *Can. J. Bot.* **82**: 1063-1073.
- Zeitlmayr, L. 1968. *Wild Mushrooms: an Illustrated Handbook*. F. Muller, London. pp. 138.
- Zhang, J., Shi, R., Li, H., Xiang, Y., Xiao, L., Hu, M., Ma, F., Ma, C.W. and Huang, Z. 2016. Antioxidant and neuroprotective effects of *Dictyophora indusiata* polysaccharide in *Caenorhabditis elegans*. *J. Ethnopharmacol.* **192**: 413-422.